

# Microarrays Denoising via Smoothing of Coefficients in Wavelet Domain

Mario Mastriani, and Alberto E. Giraldez

**Abstract**—We describe a novel method for removing noise (in wavelet domain) of unknown variance from microarrays. The method is based on a smoothing of the coefficients of the highest subbands. Specifically, we decompose the noisy microarray into wavelet subbands, apply smoothing within each highest subband, and reconstruct a microarray from the modified wavelet coefficients. This process is applied a single time, and exclusively to the first level of decomposition, i.e., in most of the cases, it is not necessary a multiresolution analysis. Denoising results compare favorably to the most of methods in use at the moment.

**Keywords**—Directional smoothing, denoising, edge preservation, microarrays, thresholding, wavelets

## I. INTRODUCTION

A microarray is affected by noise in its acquisition and processing. Microarray denoising is used to remove the additive noise while retaining as much as possible the important image features. In the recent years there has been an important amount of research on wavelet thresholding and threshold selection for bioimages denoising, e.g., microarray images [1], [2], because wavelet provides an appropriate basis for separating noisy signal from the image signal. The motivation is that as the wavelet transform is good at energy compaction, the small coefficients are more likely due to noise and large coefficient due to important signal features [3]–[5]. These small coefficients can be thresholded without affecting the significant features of the image.

In general, the results of the microarray processing combine two sample images that after further image processing, gene expression data can be produced for further analysis, such as gene clustering or identification [1], [2]. These three crucial steps, experiment, image processing and data analysis, determine the success or not of the microarray analysis. Image processing plays a potentially large impact on the subsequent analysis. In recent years, a large number of commercial tools have been developed in microarray image processing [1], [2]. The tasks of all these tools mainly focus on two major targets, namely: spot segmentation and spot intensity extraction. However, the quality of the images from the experiments is not always perfect. The gene array experiments involve a

large number of error-prone steps which lead to a high level of noise in the resulting images [1], [2]. Hence, the accuracy of the gene expressions derived from these images will largely be affected in the process.

In order to assure the accuracy of the gene expression, normally the replicated experiments and incorporated statistical methods are needed to estimate the errors [1], [2]. These methods deal mainly with measurement error, such as preparation of the sample, cross hybridization, and fluctuation of fluorescence value from gene to gene. But none deals particularly with the effect of the noise [1], [2].

In fact, the thresholding technique is the last approach based on wavelet theory to provide an enhanced approach for eliminating such noise source and ensure better gene expression. Thresholding is a simple non-linear technique, which operates on one wavelet coefficient at a time. In its basic form, each coefficient is thresholded by comparing against threshold, if the coefficient is smaller than threshold, set to zero; otherwise it is kept or modified. Replacing the small noisy coefficients by zero and inverse wavelet transform on the result may lead to reconstruction with the essential signal characteristics and with less noise. Since the work of Donoho & Johnstone [5], there has been much research on finding thresholds, however few are specifically designed for images [3], [4], [6].

Unfortunately, this technique has the following disadvantages:

- 1) it depends on the correct election of the type of thresholding, e.g., OracleShrink, VisuShrink (soft-thresholding, hard-thresholding, and semi-soft-thresholding), SureShrink, Bayesian soft thresholding, Bayesian MMSE estimation, Thresholding Neural Network (TNN), due to Zhang, NormalShrink, , etc. [3]–[7],
- 2) it depends on the correct estimation of the threshold which is arguably the most important design parameter,
- 3) it doesn't have a fine adjustment of the threshold after their calculation,
- 4) it should be applied at each level of decomposition, needing several levels, and
- 5) the specific distributions of the signal and noise may not be well matched at different scales.

Therefore, a new method without these constraints will represent an upgrade.

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## II. SMOOTHING OF COEFFICIENTS (SC) IN WAVELET DOMAIN

We decompose the noisy microarray into four wavelet subbands: Coefficients of Approximation (CA), and noisy coefficients of Diagonal Detail ( $CDD_n$ ), Vertical Detail ( $CVD_n$ ), and Horizontal Detail ( $CHD_n$ ), respectively. We apply a bidimensional smoothing within each highest subband, and reconstruct a microarray from the modified wavelet coefficients, that is to say, denoised coefficients of Diagonal Detail ( $CDD_d$ ), Vertical Detail ( $CVD_d$ ), and Horizontal Detail ( $CHD_d$ ), respectively, as shown in Fig. 1, where: DWT-2D is the Bidimensional Discrete Wavelet Transform, and IDWT-2D is the inverse of DWT-2D.

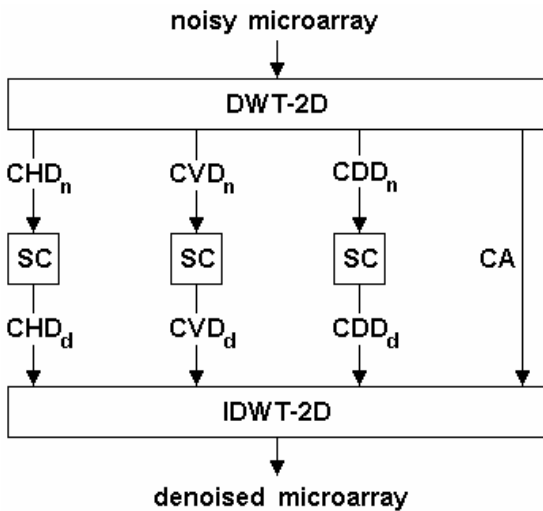


Fig. 1 Smoothing of Coefficients (SC) in wavelet domain

If we use an original microarray of  $R$ -by- $C$  pixels, then each subbands will have  $(R/2)$ -by- $(C/2)$  coefficients. The SC process is applied - in principle - a single time, and exclusively to the first level of decomposition.

On the other hand, to protect the edges from blurring while smoothing the respective coefficients of subband, an appropriate filter must be applied. The most of statistical filters have a speckle reduction approach that performs spatial filtering in a square-moving window defined as kernel, and is based on the statistical relationship between the central pixel and its surrounding pixels as shown in Fig. 2.

The size of the filter window can range from 3-by-3 to 33-by-33, with an odd number of cells in both directions. A larger filter window means that a larger area of the subband will be used for calculation and requires more computation time depending on the complexity of the filter's algorithm. If the size of filter window is too large, the important details will be lost due to over smoothing. On the other hand, if the size of the filter window is too small, speckle reduction may not be very effective. In practice, a 3-by-3 or a 7-by-7 filter window usually yields good results in the cases under study [8]-[9].

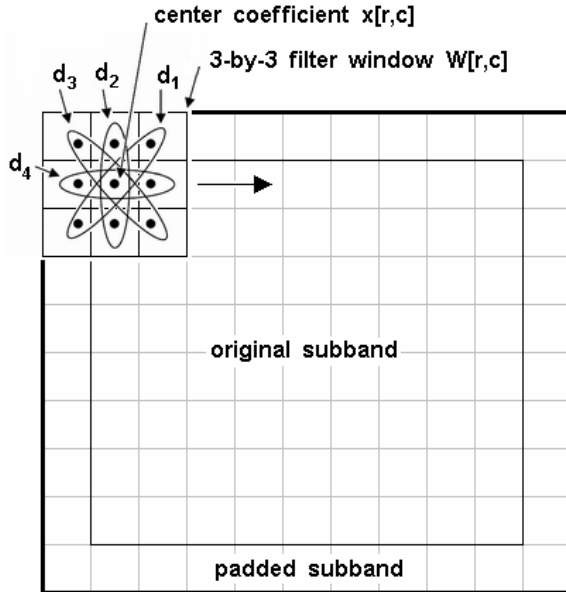


Fig. 2 3-by-3 filter window for noise smoothing over each highest subband (CHD, CVD, and CDD) using directional smoothing

For example, if the statistical filter used inside SC method (that is to say, over each highest subbands) is Directional Smoothing (DS), then, let  $x[r, c]$  denote the value of the corresponding noisy detail coefficient at location  $(r, c)$ . Let  $W[r, c]$  represent the group of coefficients contained in a filtering window with the size of 3-by-3 pixels and centered at location  $(r, c)$  of the corresponding noisy detail coefficient, as shown in Fig. 2:

$$W[r, c] = \{ x[r+p, c+q] \mid (p, q) \in [-1, 0, 1] \} \quad (1)$$

where  $p$  and  $q$  are integer indices each individually ranging from  $-1$  to  $1$ . Here,  $r$  and  $c$  are the row and the column indices, respectively, with  $2 \leq r \leq (R/2)-1$  and  $2 \leq c \leq (C/2)-1$ .

The noisy input subband is processed by sliding a 3-by-3 filtering window on the subband. The window is started from the upper-left corner of the subband and moved sideways and progressively downwards in a raster scanning fashion. Meanwhile, the directional averaging filter (selective with respect to direction) examine the average based on several directionally oriented masks, as shown in Fig. 2, and it compute averages in  $d_1, d_2, d_3$  and  $d_4$  directions  $Avg_i; i=1,2,3,4$ . Such as,  $y[r,c] = Avg_i$  where  $Avg_i$  is the one closest in amplitude to  $x(r,c)$ , i.e.  $|x(r,c) - Avg_i|$  minimum. That is to say, it has a tendency not to destroy boundaries. On the other hand, the directional analysis can also be used to check if a coefficient belongs to a directional edge (leave unchanged) or is noise (remove noise).

Finally, any used filter performs the filtering based on either local statistical data given in the filter window to determine the noise variance within the filter window, or

estimating the local noise variance of the subband under study, e.g., Median, Lee, Kuan, Gamma-Map, Enhanced Lee, Frost, Enhanced Frost [3], [8], Wiener [8], DS [8], [9] and Enhanced DS (EDS) [8].

### III. NOISE SOURCES AND ITS STATISTICAL MEASUREMENT IN MICROARRAY IMAGING

It is well known microarray technology can monitor thousand of DNA sequences in a high density array on a glass. The basic procedure for a microarray experiment is simply described as follow. Two mRNA samples are reverse-transcribed into cDNA, labeled using different fluorescent dyes (e.g., the red fluorescent dye Cy5 and the green fluorescent dye Cy3), then mixed and hybridized with the arrayed DNA sequences. After this competitive hybridization, the slides are imaged using a scanner which makes fluorescence measurement for each dye. From the differential hybridization of the two samples, the relative abundance of the spotted DNA sequences can be assessed. A schematic diagram for this process created is shown in [2].

The results of the microarray experiment are two 16-bit tagged mage files, one for each fluorescent dye. The Fig. 3(a) show an example of the mentioned microarray image. As shown in Fig. 3(b), the image is not perfect and it includes noisy sources that blur such image for further gene expression experimentation. The noise source originates from different sources during the course of experiment, such as photon noise, electronic noise, laser light reflection, dust on the slide, and so on. Hence, it is crucial to denoise the resultant image within this process.

Exciting methods to reduce the noise source include using clean glass slide and using a higher laser power rather than higher PMT voltages. However, there are not adequate for the required image qualities and an enhanced software procedure embedded within the process in a much better alternative. In this paper, we focus on the implementation of the SC method (in wavelet domain) to the denoising on microarray images [2].

Yet there are some fundamental obstacles that need clarification before the full potential of microarrays can be explored. One of the major problems in interpretation of microarray data is that different clustering techniques produce different results.

On the other hand, the assessment parameters that are used to evaluate the performance of noise reduction [10], [11] are the following ones:

#### A. Absolute Average Difference (AAD):

$$AAD = \frac{\sum_{r,c} |I(r,c) - I_d(r,c)|}{R * C} \quad (2)$$

#### B. Signal to Noise Ratio (SNR):

$$SNR = \frac{\sum_{r,c} I(r,c)^2}{\sum_{r,c} (I(r,c) - I_d(r,c))^2} \quad (3)$$

#### C. Peak Signal to Noise Ratio (PSNR):

$$PSNR = \frac{R * C * \max_{r,c} (I(r,c)^2)}{\sum_{r,c} (I(r,c) - I_d(r,c))^2} \quad (4)$$

#### D. Image Fidelity (IFy):

$$IFy = 1 - \frac{1}{SNR} \quad (5)$$

#### E. Correlation Quality (CQy):

$$CQy = \frac{\sum_{r,c} I(r,c) * I_d(r,c)}{\sum_{r,c} I(r,c)} \quad (6)$$

#### F. Structural Content (SCt):

$$SCt = \frac{\sum_{r,c} I(r,c)^2}{\sum_{r,c} I_d(r,c)^2} \quad (7)$$

Where for an image of  $\mathbf{R} * \mathbf{C}$  (rows-by-columns) pixels,  $\mathbf{r}$  means row,  $\mathbf{c}$  means column,  $\mathbf{I}$  means original image (without noise), and  $\mathbf{I}_d$  means denoised image. Such as, a lower **AAD** gives a “cleaner” image as more noise is reduced; larger **SNR** and **PSNR** indicates a smaller difference between the original (without noise) and denoised image; if **IFy** and **SCt** spread at 1, we will obtain an image  $\mathbf{I}_d$  of better quality; and a larger value of **CQy** usually corresponds to a better quantitative performance [10], [11].

On the other hand, to compare edge preservation performances of different noise reduction schemes, we adopt the Pratt's figure of merit (FOM) [9] defined by

$$FOM = \frac{1}{\max\{\hat{N}, N_{ideal}\}} \sum_{i=1}^{\hat{N}} \frac{1}{1 + d_i^2 \alpha} \quad (8)$$

Where  $\hat{N}$  and  $N_{ideal}$  are the number of detected and ideal edge pixels, respectively,  $d_i$  is the Euclidean distance between the  $i$ th detected edge pixel and the nearest ideal edge pixel, and  $\alpha$  is a constant typically set to 1/9. **FOM** ranges between 0 and 1, with unity for ideal edge detection.

#### IV. RESULTS

The simulations demonstrate that the SC technique improves the noise reduction performance to the maximum, for bioimages. Here, we present a set of experimental results using two bioimages. Such images were converted to bitmap file format for their treatment [11]. On the other hand, the statistical filters used inside SC method were DS and EDS. For statistical filters employed along, i.e., Median, Lee, Kuan, Gamma-Map, Enhanced Lee, Frost, Enhanced Frost, Wiener, DS, and EDS, we use a reduction scheme [8].

Figure 3 shows the noisy (30 %) and filtered microarray images used in the first experiment of [1], with a 274-by-274 (pixels) by 65536 (gray levels) bitmap matrix.

Table 1 summarizes the assessment parameters vs. 19 filters for Fig. 3, where En-Lee means Enhanced Lee Filter, En-Frost means Enhanced Frost Filter, ST means Soft-Thresholding, HT means Hard-Thresholding and SST means Semi-Soft-Thresholding. The assessment parameters were applied to the whole image.

Figure 4 shows the noisy (10 %) and filtered microarray images used in the second experiment of [1], with a 256-by-256 (pixels) by 65536 (gray levels) bitmap matrix.

Table 2 summarizes the assessment parameters vs. 19 filters for Figure 4.

In both cases, the bioimages were processed by using 10 statistical filters, VisuShrink with Daubechies 4 wavelet basis and 1 level of decomposition (improvements were not noticed with other basis of wavelets) [2], [3], [5], [6], [8], SureShrink, Oracle-Shrink, BayesShrink, NormalShrink, TNN [5]-[8], and SC respectively.

Figures 3 and 4 summarize the edge preservation performance of the SC technique vs. the rest of the filters with a considerably acceptable computational complexity. A 3-by-3 kernel was employed for all statistic noise filters. For TNN [7] the empirical function parameter value  $\lambda = 0.01$ .

For Lee, Enhanced Lee, Kuan, Gamma, Frost and Enhanced Frost filters the damping factor is set to 1, see [3], [8]. The quantitative results of Table 1 and 2 shows that the SC technique can eliminate noise without distorting useful image information and without destroying the important image edges. Also, in the experiment, the SC outperformed the conventional and no conventional noise reducing filters in terms of edge preservation measured by Pratt figure of merit [9]. In nearly every case in every homogeneous region, the SC produced the lowest standard deviation and was able to

preserve the mean value of the region.

The numerical results are further supported by qualitative examination, as shown in Fig. 3 and 4.

On the other hand, all filters was applied to complete image, for Figure 3 (274-by-274) pixels and Figure 4 (256-by-256) pixels, and all the filters were implemented in MATLAB® (Mathworks, Natick, MA) on a PC with an Athlon (2.4 GHz) processor.

#### V. CONCLUSIONS

In this paper we have developed a SC technique based tools for removing additive noise in microarrays. The simulations show that the SC have better performance than the most commonly used filters for microarrays (for the studied benchmark parameters) which include statistical filters, wavelets, and a version of TNN. The SC exploits the local coefficient of variations in reducing noise. The performance figures obtained by means of computer simulations reveal that the SC technique provides superior performance in comparison to the above mentioned filters in terms of smoothing uniform regions and preserving edges and features. The effectiveness of the technique encourages the possibility of using the approach in a number of ultrasound and radar applications. Besides, the method is computationally efficient and can significantly reduce the noise while preserving the resolution of the original microarray image. Considerably increased Pratt's figure of merit strongly indicates improvement in detection performance. Also, cleaner images suggest potential improvements for classification and recognition.

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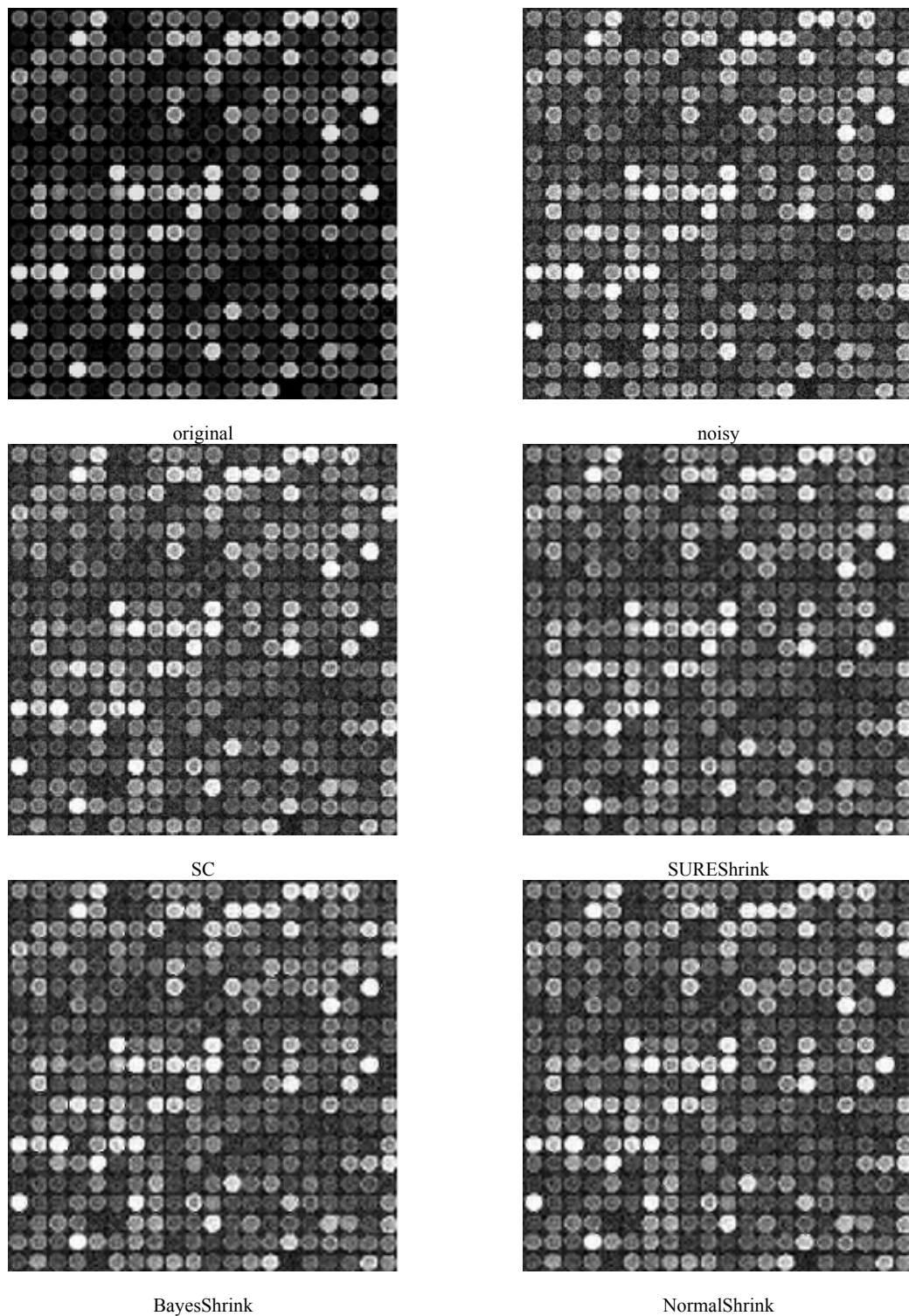
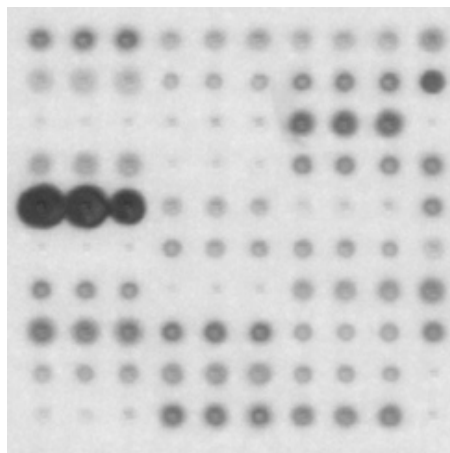
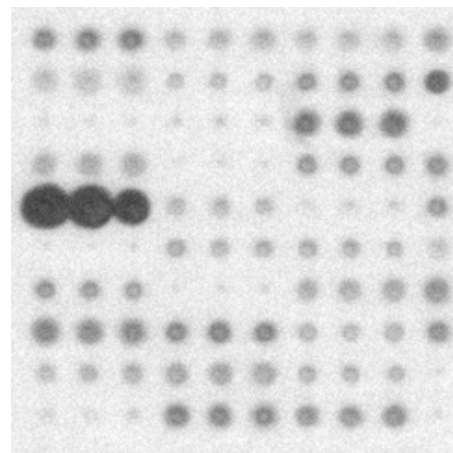


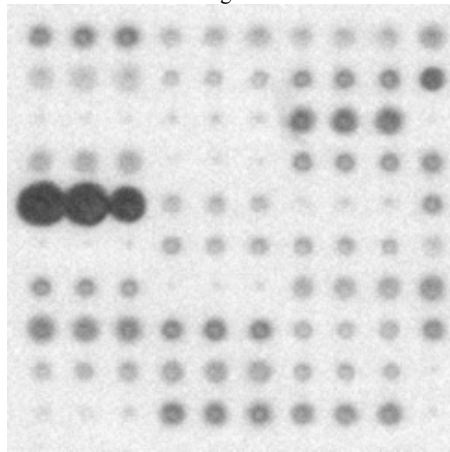
Fig. 3 Original, noisy and filtered images



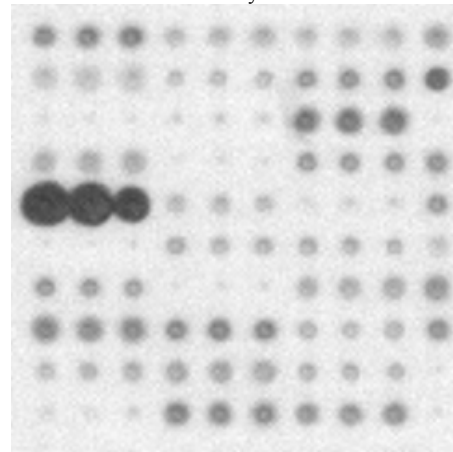
original



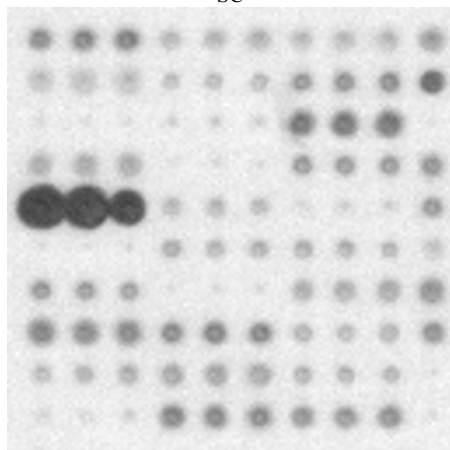
noisy



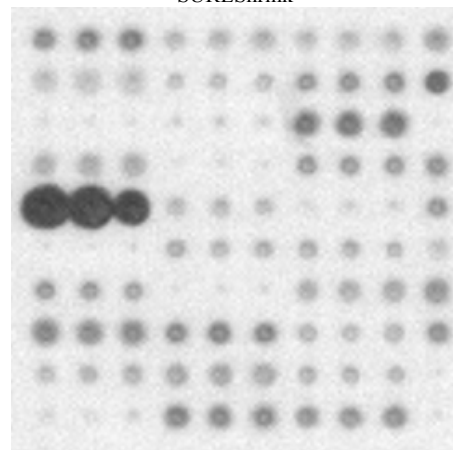
SC



SUREShrink



BayesShrink



NormalShrink

Fig. 4 Original, noisy and filtered images.

TABLE I  
ASSESSMENT PARAMETERS VS. FILTERS FOR FIGURE 3

Filter	Assessment Parameter						
	AAD	SNR	PSNR	IF	CQ	SC	FOM
En-Frost	38.2653	3.4464	33.7364	0.7109	150.7467	0.5663	0.39857
En-Lee	39.7437	3.3363	33.8373	0.7112	150.7472	0.5632	0.49876
Frost	38.4374	3.2423	33.7033	0.7106	150.5244	0.5689	0.48756
Lee	39.2427	3.4242	32.6363	0.7015	150.4141	0.5924	0.43447
Gamma	39.6252	3.1112	33.2703	0.7063	150.1918	0.5751	0.44235
Kuan	39.8224	3.1243	31.8272	0.7041	149.3121	0.5715	0.45342
Median	39.5252	3.1131	32.7916	0.6852	148.9172	0.5896	0.40704
Wiener	39.1829	3.4557	33.7033	0.7106	150.5244	0.5689	0.44236
DS	38.7332	3.4657	33.9997	0.7169	150.9898	0.5599	0.64111
EDS	38.1484	3.6969	34.1315	0.7182	151.5252	0.5612	0.64324
VisuShrink (ST)	39.1450	3.4596	33.7412	0.7109	151.1527	0.5657	0.44382
VisuShrink (HT)	38.8612	3.5283	34.4115	0.7166	151.3316	0.5666	0.44324
VisuShrink (SST)	38.1829	3.5557	34.7033	0.7196	151.9202	0.5612	0.46432
SureShrink	38.1612	3.5751	34.7193	0.7198	151.9244	0.5611	0.43322
OracleShrink	38.1189	3.6957	34.7233	0.7198	151.9844	0.5619	0.45534
BayesShrink	38.1145	3.6968	34.7237	0.7199	151.9953	0.5612	0.46329
NormalShrink	38.1098	3.6998	34.8734	0.7199	151.9983	0.5609	0.59333
TNN	38.1008	3.7157	34.8833	0.7199	151.9992	0.5600	0.65432
SC	37.7155	3.7772	36.8388	0.7353	155.4613	0.5513	0.69123

TABLE II  
ASSESSMENT PARAMETERS VS. FILTERS FOR FIGURE 4

Filter	Assessment Parameter						
	AAD	SNR	PSNR	IF	CQ	SC	FOM
En-Frost	12.4747	290.1324	363.6712	0.9830	226.4744	0.8972	0.41265
En-Lee	12.8474	290.2522	363.9321	0.9883	226.8373	0.8932	0.51986
Frost	12.1847	290.2772	363.0233	0.9828	226.3272	0.8923	0.55312
Lee	12.3733	290.2333	363.0238	0.9838	226.2822	0.8943	0.44421
Gamma	12.3830	290.8331	363.3433	0.9882	226.8383	0.8934	0.51235
Kuan	12.3833	290.8272	363.4923	0.9887	226.8381	0.8934	0.54129
Median	12.9973	289.1212	361.8374	0.9673	225.9287	0.8734	0.51286
Wiener	11.9042	290.8635	363.5568	0.9866	226.8901	0.8954	0.56413
DS	11.4572	290.9950	363.9393	0.9898	226.9723	0.8993	0.64213
EDS	11.5792	290.9998	363.9865	0.9899	226.9975	0.8993	0.64449
VisuShrink (ST)	11.9055	289.2367	361.5523	0.9761	222.7564	0.8872	0.51228
VisuShrink (HT)	11.9042	290.8673	363.5615	0.9966	226.8909	0.8976	0.56424
VisuShrink (SST)	11.7864	290.9546	363.9822	0.9975	226.8937	0.8984	0.56389
SureShrink	11.7074	291.0753	363.8343	0.9991	226.8942	0.8991	0.57432
OracleShrink	11.8436	290.9332	363.7363	0.9968	226.8963	0.8983	0.55234
BayesShrink	11.9353	290.9363	363.7361	0.9923	226.8942	0.8962	0.56328
NormalShrink	11.6875	290.9992	363.9353	0.9992	226.9021	0.8999	0.59611
TNN	11.4447	291.7243	363.9991	0.9994	226.9732	0.9002	0.62900
SC	10.9071	294.9237	383.1090	0.9992	229.8972	0.9173	0.69322