

In vitro Culture Medium Sterilization by Chemicals and Essential Oils without Autoclaving and Growth of Chrysanthemum Nodes

Wittaya Deein, Chockpisit Thepsithar, Aree Thongpukdee

Abstract—Plant tissue culture is an important *in vitro* technology applied for agricultural and industrial production. A sterile condition of culture medium is one of the main aspects. The alternative technique for medium sterilization to replace autoclaving was carried out. For sterilization of plant tissue culture medium without autoclaving, ten commercial pure essential oils and 5 disinfectants were tested. Each essential oil or disinfectant was added to a 20-mL Murashige and Skoog (MS) medium before medium was solidified in a 120-mL container, kept for 2 weeks before evaluating sterile conditions. Treated media, supplemented with essential oils or disinfectants, were compared to control medium, autoclaved at 121 degree Celsius for 15 min. Sterile conditions of MS medium were found 100% from betel oil or clove oil (18 μ L/20 mL medium), cinnamon oil (36 μ L/20 mL medium), lavender oil or holy basil oil (108 μ L/20 mL medium), and lemon oil or tea tree oil or turmeric oil (252 μ L/20 mL medium), compared to 100% sterile condition from autoclaved medium. For disinfectants, 2% iodine + 2.4% potassium iodide, 2% merbromine solution, 10% povidone-iodine, 6% sodium hypochlorite or 0.1% thimerosal at 36 μ L/20 mL medium provided 100% sterile conditions. Furthermore, growth of new shoots from chrysanthemum node explants on treated media (fresh weight, shoot length, root length and number of node) were also reported and discussed in the comparison of those on autoclaved medium.

Keywords—Chrysanthemum, disinfectants, essential oils, MS medium, sterilizing agents, sterilization of medium without autoclaving.

I. INTRODUCTION

PLANT tissue culture is a very useful technology for plant propagation. The knowledge has been provided to agriculturists worldwide. Unfortunately, most agriculturists cannot carry out plant tissue culture laboratory by themselves due to high production costs. One of the major problems is expensive equipment especially an autoclave, a sterilizing apparatus. Therefore, the development of techniques, using chemicals or plant extracts or in combinations for eradicating microorganisms causing agents of contamination, to replace the autoclaving method for establishing aseptic culture

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medium will be the best procedure for plant tissue culture.

The use of biocides, disinfectants, fungicides and bactericides such as chlorine, sodium hypochlorite, calcium hypochlorite, hydrogen peroxide, methylchloroisothiazolinone, and chemical mixtures containing methylisothiazolinone, magnesium chloride, magnesium nitrate, potassium sorbate and sodium benzoate supplemented in culture medium for preventing contamination was reported [1], [2]. Sterile culture media without autoclaving of some plants including orchids were reported using sodium hypochlorite or sodium dichloroisocyanurate [3]-[6]. The studies on plant extracts and essential oils as microorganism inhibitors were reported on betel (*Piper betle* L.) [7]-[9], cassumunar ginger (*Zingiber cassumunar* Roxb.) [10], holy basil (*Ocimum sanctum* L.) and clove (*Eugenia caryophyllata* Thunb.) [11], lavender (*Lavandula angustifolia* Mill.) [12], [13], lemon [*Citrus limon* (L.) Burm. F.] and bergamot (*C. bergamia* Risso) [14] and turmeric (*Curcuma longa* L.) [15].

This research reported effects of disinfectants and plant essential oils as sterilizing agents on sterile condition of MS medium and growth of chrysanthemum nodes on treated medium.

II. MATERIALS AND METHODS

A. Medium Used

The medium used for *in vitro* culture of chrysanthemum nodes was Murashige and Skoog (MS) medium [16] supplemented with 30 g/L sucrose and 5.5 g/L agar (Hardy Diagnostics Criterion agar, Bacteriological grade, USA). The pH of the medium was adjusted to 5.8. Each essential oil (bergamot oil, betel oil, cassumunar ginger oil, cinnamon oil, clove oil, holy basil oil, lavender oil, lemon oil, tea tree oil and turmeric oil) or disinfectant (0.1% thimerosal, 2% merbromine solution, 2% iodine + 2.4% potassium iodide, 10% povidone-iodine and 6% sodium hypochlorite) was added in a 120-mL glass jar containing 20 mL of heated culture medium in various concentrations (9 – 396 μ L). All media were kept in room temperature (about 29 \pm 2°C) for 2 weeks to investigate effects of sterilizing agents on sterile conditions of media compared to autoclaved medium. Node explants of chrysanthemum ‘Moneymaker Improved’, about 1 cm. long with 2 nodes were cultured on sterilizing agent-treated media for 5 weeks.

B. Culture Conditions

All cultures were incubated under a $24 \pm 1^\circ\text{C}$ with a 16 h photoperiod at $35 - 40 \mu\text{mole}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ provided by cool white lights.

C. Statistical Analysis

Sterile conditions of media were evaluated after 2 weeks with 20 replications. Growth of a new shoot from a chrysanthemum node, whole fresh weight (FW), shoot length, root length and number of root were collected after 5 weeks of culturing. Each treatment was replicated 10 times. The completely randomized design (CRD) was used as the experimental design and means were compared by Duncan's New Multiple Range Test at $P = 0.05$ [17].

III. RESULTS

A. Effects of Sterilizing Agents on Sterile Conditions of Treated Media

A 20-mL MS medium was treated with each sterilizing agent, kept in room temperature (about $29 \pm 2^\circ\text{C}$) for 2 weeks. For MS medium, complete sterilization (100%) of culture medium was found from medium supplemented with bergamot oil, cassumunar ginger oil, lemon oil, tea tree oil or turmeric oil at $252 \mu\text{L}/20 \text{ mL}$ medium, betel oil or clove oil at $18 \mu\text{L}/20 \text{ mL}$ medium, cinnamon oil, 2% iodine + 2.4% potassium iodide, 2% merbromin solution, 10% povidone-iodine, 6% sodium hypochlorite or 0.1% thimerosal at $36 \mu\text{L}/20 \text{ mL}$ medium and holy basil or lavender oil at $108 \mu\text{L}/20 \text{ mL}$ medium. However, 85 – 95% sterile conditions of medium were found from bergamot oil, holy basil oil or lavender oil at $36 \mu\text{L}/20 \text{ mL}$ medium, betel oil or clove oil at $9 \mu\text{L}/20 \text{ mL}$ medium, cassumunar ginger oil at $18 \mu\text{L}/20 \text{ mL}$ medium, lemon oil at $180 \mu\text{L}/20 \text{ mL}$ medium and tea tree oil at $108 \mu\text{L}/20 \text{ mL}$ medium (Table I).

B. Effects of Sterilizing Agent-Treated Media on Growth of Chrysanthemum Nodes

Medium supplemented with disinfectants (36 and $108 \mu\text{L}/20 \text{ mL}$ medium) and essential oils (the concentrations that gave 85 – 100% sterile conditions of medium) were chosen for culturing chrysanthemum nodes. It was found that there was no growth and explants died on media added with bergamot oil, cassumunar ginger oil, holy basil oil, tea tree oil, clove oil ($18 \mu\text{L}$), lavender oil ($108 \mu\text{L}$), 2% merbromin solution and 0.1% thimerosal (Table II).

For essential oils, betel oil at $9 \mu\text{L}/20 \text{ mL}$ medium and lemon oil at $180 \mu\text{L}/20 \text{ mL}$ medium (providing 90% sterile conditions of medium) gave 0.252 and 0.180 g. FW, 2.11 and 1.89 cm. shoot length, 5.12 and 8.19 cm. root length with 5.5 and 5.3 nodes, respectively (Table II; Figs. 1 (g) and (i)). Other essential oils (providing 100% sterile conditions) gave 0.060 – 0.105 g. FW, 0.10 – 0.92 cm. shoot length, 0.00 – 2.66 cm. root length and 1.0 – 4.5 nodes (Table II; Figs. 1 (h) and (j)). Some essential oils, lavender oil and clove oil (providing 85 and 95% sterile conditions) gave 0.155 and 0.115 g. FW, 1.72 and 0.70 cm. shoot length, 6.89 and 2.29 cm. root length with 7.1 and 3.5 nodes, respectively (Table II; Figs. 1 (k) and (l)).

TABLE I
PERCENTAGE OF STERILE MS MEDIUM AFTER TREATED WITH DIFFERENT ESSENTIAL OILS AND DISINFECTANTS FOR 2 WEEKS

Treatments	Sterile conditions (%) ¹							
	Concentration (μL) in a 20-mL MS medium							
	9	18	36	108	180	252	324	396
Autoclaved	100							
Bergamot oil	35	65	85	85	85	100	-	-
Betel oil	90	100	100	100	100	100	-	-
Cassumunar ginger oil	75	85	90	90	95	100	-	-
Cinnamon oil	35	40	100	100	100	100	-	-
Clove oil	95	100	100	100	100	100	-	-
Holy basil oil	20	65	95	100	100	100	-	-
Lavender oil	20	40	85	100	100	100	-	-
Lemon oil	45	45	60	80	90	100	-	-
Tea tree oil	35	35	45	95	95	100	-	-
Turmeric oil	10	20	45	60	75	100	-	-
0.1% Thimerosal	-	-	100	100	100	100	100	100
2% Merbromin solution	-	-	100	100	100	100	100	100
2% Iodine+2.4% KI	-	-	100	100	100	100	100	100
6% NaOCl	-	-	100	100	100	100	100	100
10% povidone-iodine	-	-	100	100	100	100	100	100

N = 20

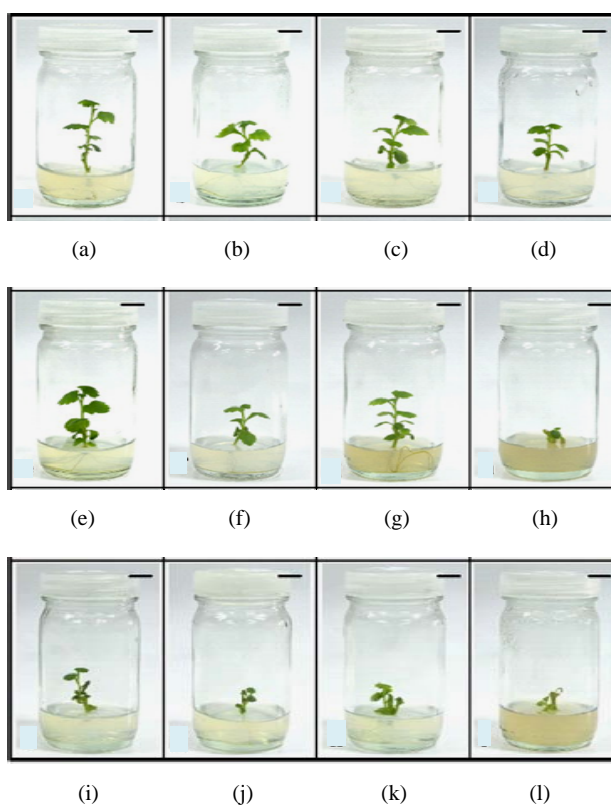


Fig. 1 Growth of a new shoot from chrysanthemum node explant on sterilizing agent treated 20-mL MS medium cultured for 5 weeks (bar = 1 cm.) (a) = autoclaved medium (control); (b) = $36 \mu\text{L}$ of 6% sodium hypochlorite; (c) = $36 \mu\text{L}$ of 2% iodine + 2.4% potassium iodide; (d) = $108 \mu\text{L}$ of 2% iodine + 2.4% potassium iodide; (e) = $36 \mu\text{L}$ of 10% povidone-iodine; (f) = $108 \mu\text{L}$ of 10% povidone-iodine; (g) = $9 \mu\text{L}$ of betel oil; (h) = $18 \mu\text{L}$ of betel oil; (i) = $180 \mu\text{L}$ of lemon oil; (j) = $252 \mu\text{L}$ of lemon oil; (k) = $36 \mu\text{L}$ of lavender oil; (l) = $9 \mu\text{L}$ of clove oil

TABLE II

GROWTH OF A NEW SHOOT FROM A CHRYSANTHEMUM 'MONEYMAKER IMPROVED' NODE EXPLANT ON TREATED MS MEDIUM AFTER CULTURING FOR 5 WEEKS

Treatment ($\mu\text{L}/20\text{ mL medium}$)		Growth of chrysanthemum node ²			
Sterilizing agent	amount (μL) ¹	Fresh weight (g)	Shoot length (cm)	Root length (cm)	No. of node
Autoclaved	-	0.342±0.003 a	2.88±0.03 a	7.41±0.02 f	5.5±0.2 c
Bergamot oil	252	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h
Betel oil	9	0.252±0.002 d	2.11±0.03 d	5.12±0.02 i	5.5±0.2 c
Betel oil	18	0.105±0.002 i	0.92±0.02 i	2.66±0.05 j	2.5±0.2 f
Cassumunar ginger oil	95	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h
Cinnamon oil	36	0.045±0.002 l	0.10±0.00 m	0.00±0.00 n	1.0±0.0 g
Clove oil	9	0.115±0.002 k	0.70±0.03 k	2.29±0.03 l	3.5±0.2 e
Lavender oil	36	0.155±0.002 g	1.72±0.03 f	6.89±0.02 g	7.1±0.2 a
Lemon oil	180	0.180±0.001 ef	1.89±0.02 e	8.19±0.03 e	5.3±0.2 c
Lemon oil	252	0.075±0.002 j	0.82±0.03 j	2.47±0.02 k	4.5±0.2 d
Holy basil oil	108	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h
Tea tree oil	108	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h
Turmeric oil	252	0.060±0.000 k	0.21±0.02 l	0.39±0.03 m	3.6±0.2 e
2% iodine+2.4% KI	36	0.306±0.002 b	2.22±0.03 c	11.03±0.04 c	7.2±0.2 a
2% iodine+2.4% KI	108	0.176±0.002 f	1.62±0.03 g	13.51±0.02 b	6.1±0.2 b
2% merbromine solution	36	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h
10% povidone-iodine	36	0.345±0.002 a	2.52±0.03 b	9.12±0.03 d	6.9±0.2 a
10% povidone-iodine	108	0.185±0.002 e	1.52±0.03 h	17.12±0.04 a	5.5±0.2 c
6% NaOCl	36	0.290±0.002 c	1.90±0.03 e	6.13±0.03 h	5.9±0.3 cd
0.1% thimerosal	36	0.000±0.000 m	0.00±0.00 n	0.00±0.00 n	0.0±0.0 h

¹ amount of essential oils or disinfectants (μL) added in 20-mL MS medium per container.² Values are mean \pm SE (n = 10). Means followed by the same letters within the same column are not significantly different at $P = 0.05$ by Duncan's new multiple range test.

IV. DISCUSSIONS

In the experiment, 10% povidone-iodine or 2% iodine + 2.4% potassium iodide or 6% sodium hypochlorite at 36 $\mu\text{L}/20\text{ mL}$ medium were promising to use as sterilizing agents in solid MS medium. These disinfectants provided completely sterile condition of MS medium. The disinfectant-treated medium could be used for culturing chrysanthemum node explants. The results were similar to the reports of Teixeira et al. [4], Yanagawa et al. [5] and Chansean and Syoichi [6]. Culture media, for wild orchid seed germination, *Cymbidium* and *Phalaenopsis* micropropagation, were sterilized by adding sodium hypochlorite solution at the appropriate concentrations of 0.005% active chlorine [5], [6]. Teixeira et al. [4] reported that active chlorine at the concentrations of 0.0003% or 0.0005% provided complete sterilization of culture medium for pineapple micro-propagation.

Plant extracts and plant essential oils showed antimicrobial activity. Fungicidal efficacy against *Rhizoctonia solani*, *Aspergillus flavus* and *Fusarium verticillioides* was found from betel [8], [9]. Antibacterial activity against some food-borne pathogens, gram negative and/or gram positive bacteria was found from betel [7], cassumunar ginger [10], clove [11], lavender [12], [13], lemon and bergamot [14] and turmeric [15]. For essential oils, growth of a new shoot from a chrysanthemum node explant was observed from MS medium added with betel oil at 9 $\mu\text{L}/20\text{ mL}$ medium or lemon oil at 180 $\mu\text{L}/20\text{ mL}$ medium. However, the concentrations of these two essential oils provided only 90% sterile condition of MS medium. Very poor growth or no growth of chrysanthemum

nodes was found from essential oils at the concentrations that provided completely sterile condition of MS medium.

This was the first report on using plant essential oils as sterilizing agents to eliminate microorganisms in MS medium to obtain sterile condition without autoclaving. Further experiments are needed to establish appropriate concentrations of single essential oil or various combinations as sterilizing agents in MS medium used for plant tissue culture.

V. CONCLUSION

Disinfectants, 2% iodine + 2.4% potassium iodide, 10% povidone-iodine or 6% sodium hypochlorite at the appropriate concentrations (36 $\mu\text{L}/20\text{ mL}$ medium), were effective for eradicate microorganisms, causal agents of *in vitro* contamination, and provided completely sterile condition of solid MS medium without autoclaving. MS medium treated with these sterilizing agents can be used for culturing chrysanthemum 'MoneyMaker Improved' nodes. For essential oils, growth of chrysanthemum nodes was found only on MS medium added with betel oil (9 $\mu\text{L}/20\text{ mL}$ medium) and lemon oil (180 $\mu\text{L}/20\text{ mL}$ medium) that provided 90% sterile condition of MS medium.

ACKNOWLEDGMENT

This research was financial supported by Thailand Toray Science Foundation (TTSF) and Faculty of Science, Silpakorn University, Thailand.

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