

# Hypothesis of a Holistic Treatment of Cancer: Crab Method

Devasis Ghosh

**Abstract**—The main hindrance to total cure of cancer is a) the failure to control continued production of cancer cells, b) its sustenance and c) its metastasis. This review study has tried to address this issue of total cancer cure in a more innovative way. A 10-pronged “CRAB METHOD”, a novel holistic scientific approach of Cancer treatment has been hypothesized in this paper. Apart from available Chemotherapy, Radiotherapy and Oncosurgery, (which shall not be discussed here), seven other points of interference and treatment has been suggested, i.e. 1. Efficient stress management. 2. Dampening of ATF3 expression. 3. Selective inhibition of Platelet Activity. 4. Modulation of serotonin production, metabolism and 5HT receptor antagonism. 5. Auxin, its anti-proliferative potential and its modulation. 6. Melatonin supplementation because of its oncogenic properties. 7. HDAC Inhibitors especially valproic acid use due to its apoptotic role in many cancers. If all the above stated seven steps are thoroughly taken care of at the time of initial diagnosis of cancer along with the available treatment modalities of Chemotherapy, Radiotherapy and Oncosurgery, then perhaps, the morbidity and mortality rate of cancer may be greatly reduced.

**Keywords**—ATF3 dampening, auxin modulation, cancer, platelet activation, serotonin, stress, valproic acid.

## I. INTRODUCTION

THIS review study has hypothesized a holistic 10 pronged “Crab Method” of treatment of cancer. It has tried to take into consideration the three main factors for failure of Cancer treatment, namely a) continued production of malignant cells, b) the ability of malignant cells to sustain for prolonged periods and c) distant spread of cancer cells.

With the intention of improving the mortality and morbidity of the dreaded disease, Cancer (Latin word for Crab having 10 legs), seven different points of interference has been suggested (other than available Chemotherapy, Radiotherapy and Oncosurgery). They are namely: 1) Mental Stress, 2) ATF3 gene, 3) Platelet Activation, 4) Serotonin, 5) Auxin, 6) Melatonin, and 7) HDAC Inhibitors. The causal relation with cancer of these above factors and the probable mode of interference is the main focus of this study.

### A. Stress and Cancer

Recent studies have found a strong association of both ‘obesity’ and ‘mental stress’ with the increasing incidence of cancers at various sites and their adverse effects on the morbidity and mortality [1], [2].

When mice were fed on high fat diet to induce obesity, there was a marked increase in the progression of melanoma

[3].

Another mouse model study has found that stress induced  $\beta$ -adrenergic activation increased macrophage infiltration into primary tumor cells, thereby inducing pro-metastatic gene expression in primary breast cancer. This has led to 30-fold increase in metastasis to distant tissues. Treatment with  $\beta$ -antagonist drugs has been effective in inhibiting metastasis. Moreover, inhibition of macrophage infiltration by Colony stimulating factor receptor (CSF-1R) inhibitor, GW2580, has also been found to be effective in retarding secondary spread [4].

### B. ATF3 and Cancer

Amongst the mammalian ATF/CREB family of transcription factors in the macrophages, Activating Transcription Factor 3 (ATF3) is a transcriptional repressor involved in cellular responses to extracellular signals [5], [6].

The mRNA level of ATF3 gene is greatly increased when the cells are exposed to stress signals, both in animal experiments, and in cultured cells [7], including many of those encountered by cancer cells (tumor microenvironment) e.g. Prostate carcinoma, Breast carcinoma, Hodgkin lymphoma [8].

ATF3 then regulates the expression of a variety of genes, in inflammation (mediated by immunoglobulin & immune complexes) which has got a significant role to play in cancer development and progression [9]-[12].

A recent study has found that ATF3 gene expressed in non-cancer host stromal mononuclear cells, but not cancer epithelial cells, leads to worst outcome and may act as an independent predictor for breast cancer death thereby signifying the importance of host stress response. Supporting data from mouse models has shown less efficient breast cancer metastasis in Atf3-deficient mice than in WT (Atf3<sup>+/+</sup>) mice. So, dampening ATF3 expression in the host may be a potential therapeutic approach [13].

Chemotherapeutic drugs presently in use have strangely both anti-cancer properties and a pro cancer effect by increasing chemo-resistance and cancer metastasis [17]. The chemotherapeutic agents which also acts as stressors, including Paclitaxel [PTX] [14], cisplatin [15], and doxorubicin [16] induces ATF3 gene in the tumor microenvironment (non-cancer host cells) which in turn plays an important role in producing this paradoxical effect.

Actually, the cellular stress response which evolved to promote tissue repair, has preferred to help cancer cell survival and progression [18], [19].

As the ability of PTX to exacerbate metastasis was completely abolished in ATF3 deficiency in host, rationally

Devasis Ghosh (Dr.) is presently working as a Consultant Physician at Belle Vue Clinic, Kolkata: 700 017, India (phone: +9198303 82967, e-mail: devasisghosh@hotmail.com).

ATF3 dampening may improve the efficacy of chemotherapy.

### C. Platelet Activation

Platelet activity is initiated and increased by physical, and more so by emotional stress, as well as stress-related psychiatric or somatic disorders. This is mediated by the activating properties of epinephrine and nor-epinephrine on the platelet.

There are increased levels of platelet 5-HT in paranoid schizophrenia and increased platelet 5-HT<sub>2A</sub> receptor density in depression [20]-[29].

The activation of platelets is followed by platelet bridging and platelet aggregation with the binding of fibrinogen to its major receptor, active form of glycoprotein receptor GPIIb/IIIa [25]. The activated platelets also adhere to Lymphocytes mainly via platelet P-selectin and lymphocyte P-selectin glycoprotein ligand-1 (PSGL-1) and sialyl saccharides, thus forming platelet-leukocyte aggregates, PLAs. Therefore, PLAs are suggested to be a very sensitive marker of platelet activation in vivo [30]-[32]. A study on the correlation of PLA and acute mental stress showed peak level at 30 min, returning to base level at 75 min [33], [34].

Activated platelets have quite a significant role in both

- a) Tumor angiogenesis
- b) Tumor metastasis.

#### 1. Tumor Angiogenesis Role

Activated platelets can secrete pro-angiogenic growth factors like platelet-derived growth factor (PDGF) and vascular endothelial growth factor (VEGF), protease like MMP9; phospholipids and other microparticles to promote angiogenesis. Activated platelets can also directly bind to Endothelial cells (EC) to support angiogenesis [35], [36]. The platelet activation and coagulation product fibrin, commonly exist in tumors, along which ECs can survive and migrate to form new blood vessels.

Activated platelets can recruit bone marrow-derived cells to the site of neovascularization by secretion from  $\alpha$ -granules, which suggests a role of platelets as communicators between hypoxic tissue and the bone marrow [37]. Platelet derived factors support tumor angiogenesis by protecting the integrity of the angiogenic and inflamed tumor vessels. [36], [38], [39].

Tumor cells secrete a Vascular permeability factor (VPF), which shows a significant homology with PDGF and VEGF. VPF has been found contribute to tumor angiogenesis. Moreover VPF also induces an increase of von Willebrand factor (vWF) which initiates platelet aggregation [40].

#### 2. Tumor Metastasis Role

Tumor cells with platelet activation capacity can form more metastasis in mice with xenografted tumors [41]. Metastasis was reduced grossly in cases of thrombocytopenia in several mouse models [42], [43].

Below three mechanisms for the metastasis supporting role of platelets are described.

1. Tumor cells are protected from shear stress and immune cell attack in the blood circulation by the platelets after intravasation by acting as a physical guard to help them

escape immune elimination [44]. Platelets may also inhibit Natural Killer (NK) cell cytotoxicity via platelet-derived Transforming growth factor beta TGF- $\beta$  [45] or other secreted factors released upon activation [46].

2. Activated platelets encourage tumor cells to roll and tether on the vessel wall, which are necessary for the extravasation. Selectins on the platelet surface could promote tumor cell (expressing selectin ligands) adherence to the endothelium transiently [47]. Moreover, the  $\beta$ 3 integrin-mediated binding to activated platelets is an efficient mechanism for melanoma cell arrest under flow, and this may contribute to the role of platelets in metastasis [48].
3. Platelet secretes prometastatic factors and matrix degrading enzymes to facilitate metastasis. From the  $\alpha$ -granules of the platelets, VEGF-A, Epidermal growth factor (EGF), PDGF and TGF- $\beta$  and serotonin and histamine from dense granules of the platelets are released which affect the vascular permeability and enhance metastasis [49].

Direct contact of the tumor cells with platelets together with Platelet-derived TGF- $\beta$ ; activate the TGF- $\beta$ /Smad and nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathways in cancer cells, resulting in their transition to an invasive mesenchymal-like phenotype and increased metastasis in vivo. Ablation of TGF $\beta$ 1 expression solely in platelets and inhibition of NF- $\kappa$ B signaling in cancer cells protects against lung metastasis in vivo [50].

In tumor cells, NF- $\kappa$ B is active either due to mutations in genes encoding the NF- $\kappa$ B transcription factors themselves or in genes that control NF- $\kappa$ B activity (such as I $\kappa$ B genes); in addition, some tumor cells secrete factors that cause NF- $\kappa$ B to become active.

Blocking NF- $\kappa$ B can cause tumor cells to stop proliferating, to die, or to become more sensitive to the action of anti-tumor agents. Thus, NF- $\kappa$ B is a target for anti-cancer therapy [51]. Disulfiram, olmesartan and dithiocarbamates can NF- $\kappa$ B signaling cascade [52].

#### LPA & MMP

Recently, the importance of platelets in the progression of malignant tumors has been studied [53], [54]. LPA (lysophosphatidic acid), which is a simple lipid with mitogenic properties is rapidly released by activated platelets [55], [56]. Moreover, LPA has been found to be useful as a diagnostic and prognostic biomarker of ovarian cancer [57]. In the initiation and progression of several cancers, such as colon, ovarian, prostate, breast, melanoma and thyroid, LPA is involved [58], [59]. The effects of LPA are mediated by at least six different G protein-coupled receptors [LPA1-6] [60]. Selective blockage of LPA1 inhibits cancer cell proliferation and invasion. [61]-[63]. The activity of matrix metalloproteinases, MMP2, MMP7 and MMP9 in cancer cells are up regulated by LPA [64]-[67].

The MMPs are enzymes that have role in cancer progression. MMPs especially MMP-9, have a critical role in certain aspects of tumor metastasis such as tumor-induced angiogenesis, tumor invasion, and establishment of metastatic

foci at the secondary site [68], [69]. A study in a mouse model has shown that induced thrombocytopenia leads to destabilization of tumor vessels with intratumoral hemorrhage, thereby reducing tumor cell proliferation and increased necrosis [70].

Recent studies have shown that in cancer patients, selective inhibition of platelet activity by low molecular weight heparins (LMWHs) not only reduces risk of embolism, but also effectively inhibits metastasis by suppression of angiogenesis and thus improving their survival rates [71]-[74]. So, the process of platelet activation may be tried as a therapeutic intervention. Lung metastasis and inhibition of TCIPA (tumor cell-induced platelet aggregation) can be greatly reduced by the oral GPIIb/IIIa antagonist, XV454, in a mouse model [75].

#### *D. Serotonin and Cancer*

Psychological stress in conscious rats has been found to markedly increase the levels of extracellular 5-HT levels in basolateral amygdaloid nucleus and the prefrontal cortex due to activation of serotonergic neurons in these brain regions [76]. This study also suggests that there is a relationship between anxiety and 5-HT release in the prefrontal cortex [77].

An overview of 30 years of research on stress and 5-HT, indeed favors the hypothesis that numerous components of central serotonergic systems are sensitive to stressors [78]. Numerous stressors increase 5-HT synthesis/turnover [79].

A recent study demonstrates that FOXO3a functions as a growth factor in serum-deprived HCC (Hepatocellular carcinoma) cells and serotonin promotes the proliferation of serum-deprived HCC cells via up-regulation of FOXO3a, in the presence of sufficient levels of the serotonin receptor 5-HT<sub>2B</sub>R. Therefore, drugs targeting the serotonin-5-HT<sub>2B</sub>R-FOXO3a pathway may provide a novel target for anticancer therapy [80].

Another study provides evidence that serotonin is involved in tumor growth of hepatocellular cancer by activating downstream targets of mTOR, and therefore, serotonin-related pathways might represent a new treatment strategy [81].

In hormone refractory PC3 prostate cancer cells, 5-HT<sub>1A</sub> and to a greater extent 5-HT<sub>1B</sub> antagonists significantly inhibit growth and induce apoptosis. This effect is most likely mediated via 5-HT<sub>1A</sub> and 1B receptors. Therefore, the results imply that 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor antagonists may act as potential antineoplastic agents [82].

A study addresses the role of 5-HT in Erk1/2 and Akt activation in prostate cancer cells, which proves towards neuroendocrine factors facilitating progression and migration of prostatic cancer cells in an androgen-deficient environment. The action of 5-HT was inhibited to varying degrees by inhibitors of MAPK and PI3K, as well as by a 5-HT receptor subtype 1A antagonist. The data presented in that study also identifies 5-HT receptors as a novel target in castration-resistant PC [83].

The proliferative effect of serotonin on cholangiocarcinoma growth and the inhibition of serotonin production effectively

inhibits tumor growth has been shown in a recent study. Furthermore, it has also been found that inhibition of the serotonin receptors 5HT<sub>1A</sub>, 5HT<sub>2A</sub>, 5HT<sub>2B</sub>, 5HT<sub>4</sub> and 5HT<sub>6</sub> effectively blocked the growth promoting effects of serotonin.

The major findings of one study relate to the dysregulation of serotonin metabolism in cholangiocarcinoma. A few significant findings in that study are as follows:

- Expression of the enzyme responsible for serotonin synthesis in the gastrointestinal tract, TPH1, is up regulated in cholangiocarcinoma;
- The enzyme responsible for serotonin degradation, MAO A, is markedly decreased in cholangiocarcinoma samples;
- That this results in an overall increase in serotonin secretion from cholangiocarcinoma cells and in the bile from cholangiocarcinoma patients.

Therefore, agents that suppress serotonin production may be very much useful in the treatment of cholangiocarcinoma [84].

Serotonin has an important role in tumor growth, especially colon cancer, by regulating angiogenesis by reducing the expression of matrix metalloproteinase 12 (MMP-12) in tumor-infiltrating macrophages. This leads to lower levels of 'angiostatin' which is an endogenous inhibitor of angiogenesis. Hence, serotonin might represent a novel target for the prevention and treatment of colon cancer [85].

MMP-12 cleaves plasminogen into angiostatin, which suppresses angiogenesis in solid tumors. Enhanced transcription and activation of MMP-12 observed in tumors of serotonin-deficient mice entailed higher levels of circulating angiostatin, causing a reduction of tumor vascularity, enhanced hypoxia, and consequently, tumor necrosis [86], [87]. Serotonin-dependent effects were reproducible in Lewis lung cancer, in accordance with previous studies showing supra-normal serotonin levels to enhance lung cancer proliferation [88], [89] and macrophage-derived MMP-12 to regulate Lewis lung cancer growth [86], [87].

#### *E. Auxin and Cancer*

Auxins (Indole-3-acetic acid), a plant hormone essential for plant body growth and development interacts with its receptor TIR1 (Transport inhibition response 1), which is similar to human ubiquitin ligase enzymes (SKP2). Auxin helps TIR1 to bind to its peptide substrate tightly thus may have an important role to play in human cancer because many types of cancer are caused by dysregulation of ubiquitination (the first breast cancer susceptibility gene Brcal is a ubiquitin ligase) [91], [92]. Also serotonin in plants which regulates root development acts as a natural auxin inhibitor [90].

Either blocked degradation of oncogenic proteins/ growth-enhancing factors or accelerated degradation of growth-suppressing proteins may disrupt the pathways controlling cell cycle progression, cell death, or survival, leading to cancer development [93], [94].

Ubiquitination and the ubiquitin-mediated proteolysis play an important role in tumorigenesis and cell growth. A powerful approach for cancer treatment would be to target the components involved in these processes. The first proteasome

inhibitor for clinical use in human cancers is Bortezomib [95].

The F-box protein SKP2 (S-phase kinase-associated protein 2) forms a complex with CUL1, SKP1, and a RING finger protein RBX1, together termed SCF<sup>SKP2</sup> [96]. Several important cell cycle regulators, including p27<sup>KIP1</sup> and p21<sup>CIP1</sup> undergo ubiquitination by SKP2 [97]-[99]. SKP2 also plays a critical role in EGFR-mediated AKT ubiquitination and membrane recruitment [100].

The oncogenic potential of SKP2 was suggested by its over expression in a variety of human cancers [101], [102]; importantly, this over expression of SKP2 showed an inverse relationship with p27KIP1 [103], [104]. The protein levels of SKP2 could indicate the prognosis, inversely proportional to survival rate of patients [100], [103], [105].

Given the importance of SKP2 in regulating degradation of tumour suppressors and its clear oncogenic potential, inhibiting SKP2 may represent a unique opportunity for the treatment of different types of tumours.

A study focused on Auxin alone and its antiproliferative potential, with emphasis on modulation of the cell cycle, of natural (IAA) and synthetic (2,4-D) Auxin, showed cytostatic effects on selected human tumor cell lines, induce strong G1 arrest, along with a drastic decrease in the percentage of S-phase cells in MCF-7 cell line. This phenomenon demonstrates that Auxins may have novel, unexploited antitumor potential [106].

#### *F. Melatonin and Cancer*

Melatonin, (N-acetyl-5-methoxytryptamine) is an indole amine secreted by the pineal gland, is an oncostatic agent. It has got antioxidative [107]-[112], anti-inflammatory and anti-tumor activities [113]-[120]. It also has the capability of modulating several signal transduction pathways associated with cell survival, proliferation, apoptosis and invasion [121]-[125].

Additionally, melatonin inhibits the growth of a variety of cancers: lung [126]-[128], breast [129]-[133] prostate [134]-[137], liver [138], [139], colon [140], [141].

Anti-cancer property of melatonin is related to its different qualities e.g. anti-proliferation [126], induction of apoptosis [126]-[128] inhibition of invasion and metastasis [142], [143], anti-angiogenesis [129], [144], and enhancement of immune modulation [145], regulation of the estrogen receptor expression and trans-activation, modulation of the enzymes involved in the local synthesis of estrogens [166]. Melatonin increases the efficacy and reduces the side effects of both radio-therapy and chemotherapy [146], [147].

Melatonin, through increasing adhesion by elevating E-cadherin and  $\beta$ 1-integrin expression or modulating microfilament, can inhibit tumor invasion [148]-[150], and decreasing matrix metalloproteinases (MMPs) production [151].

The effect of melatonin on the migration of human lung adenocarcinoma A549 cells was observed and it was found that there is an association between JNK/MAPK [c-jun-N-terminal kinases (JNK)/ mitogen activated protein kinase (MAPK)], pathway and the expression of tight junction (TJ)

related proteins occluding, myosin light-chain kinase (MLCK), osteopontin (OPN). The melatonin may inhibit A549 cell proliferation and play an important role in the inhibition of tumor progression [152].

Melatonin has been found to have a synergistic effect along with Cisplatin in human cervical cancer cells [153], in high concentrations has a pro-apoptotic effect on pancreatic carcinoma cells [154], [161], renal cancer cells [155], [160] and in the treatment of neuroblastoma [156]. It has also anti cancer effects on gastro intestinal cancer [157], in breast cancer [163]. Melatonin, though inhibits apoptotic processes in normal cells, modulates autophagy and activates the intrinsic and/ or the extrinsic apoptotic pathway in cancer cells [164] and is helpful even in tumor models unresponsive to melatonin alone, by amplifying significantly the cytostatic and the cytotoxic effects of other conventional anti cancer drugs [158], [159], [162], [165], [167].

A recent study demonstrated that melatonin supplementation down-regulated, Proliferating-cell nuclear antigen (PCNA) which is a molecular marker for proliferation and reduced the viability in both lung cancer A549 and PC9 cells [127]. Melatonin as a co-treatment with conventional cancer therapies would improve the wellbeing of the patients [168].

Besides these oncostatic properties, melatonin deserves to be considered in the treatment of cancer (doses in the 100 – 500 mg/day range) for two other reasons. First, because its hypnotic-chronobiotic properties, melatonin can effectively be used for sleep disturbances, a major co-morbidity in cancer. Second, because melatonin's anxiolytic and antidepressant effects, it has a possible application in two other major co-morbidities seen in cancer patients, i.e. depression and anxiety [169].

Melatonin acts as a proteasome inhibitor [160], which has an anti-proliferative action on human breast cancer cells. Melatonin specifically inhibits estrogen induced transcription mediated by ER $\alpha$  (estrogen receptor alpha) at the ERE (estrogen responsive element) and AP1 (Activator protein 1) gene promoters [92].

#### *G. HDAC Inhibitors Specially Valproic Acid and Cancer*

Acetylation of Histones plays a key role in epigenetic regulation of gene expression in carcinogenesis. Histone deacetylases (HDAC) inhibitors induce cancer cell cycle arrest, differentiation and cell death, reduce angiogenesis and modulate immune response, and so may be used as an anti-cancer drug in combination with other anti-cancer drugs and/ or radiotherapy.

HDAC inhibitors have been approved; Vorinostat for cutaneous T-cell lymphoma (CTCL), Belinostat for peripheral T cell lymphoma (PTCL), and Panobinostat for therapy of multiple myeloma [170].

The anticancer effects of HDAC inhibitors are different and depend on a type of cancer and dose used e.g. Valproic acid (VPA) inhibit the invasiveness in bladder cancer [171].

## Mechanisms:

- a) HDAC inhibitors induces cell cycle arrest by the increased expression of cell cycle genes such as CDKN1A (Cyclin dependent kinase inhibitor p21) [172]-[174].
- b) Apoptosis in tumor cells is induced by the HDAC inhibitors by regulation of pro-apoptotic and anti-apoptotic genes [175]-[177]. The mechanisms include activation of both extrinsic and intrinsic apoptotic pathways.
- c) Autophagy induction in apoptosis-resistant cancer cells is an important feature of HDACI. It works through several signaling pathways e.g. downregulation of AKT-mTOR signaling [178], VPA in prostate cancer cells [179].
- d) Anti-cancer effect of HDACI, is also due to its capability to alter long non-coding RNA (lncRNA) expression e.g. abexinostat in breast cancer cells [180].
- e) Activation of some of protein kinases i.e. ERK (which modulate biological processes like cell growth, differentiation and apoptosis) is done by the HDACI [181].
- f) Anti angiogenic effect of VPA is done by enhancing production of the anti-angiogenic proteins thrombospondin-1 and activin A via downregulation of pro-angiogenic factors such as the basic fibroblast growth factor (bFGF) [182].
- g) HDACI induces modulation of immune response and enhances the functions of NK cells and CD8 T cells [183].

VPA inhibits the growth of pancreatic and colon, and oral squamous cell cancer cell growth by down-regulation of  $\beta$ -amyloid precursor protein (APP) [184].

VPA when administered even in lower doses to prostate cancer reduces the net proliferation rate both in androgen receptor-positive and androgen receptor-negative prostate cancer cells. This is due to the increased caspase-2 and caspase-3 activation [185]. Moreover, chronic VPA treatment results in statistically significant reduction of tumor growth and volume in vivo. This enhanced activity results from capturing the resistant quiescent cells.

It is therefore concluded that acute treatment has nominal effects on prostate cancer cell survival and proliferation, but chronic VPA results in profound decreases in proliferation, independently of androgen regulation [185]

Monoamine oxidases (MAOs) A and B are mitochondrial isoenzymes which catalyze the oxidative deamination of dietary amines and monoamine neurotransmitters, such as serotonin, norepinephrine, dopamine. Dysfunction of MAO A leads to abnormal levels of these neurotransmitters resulting in many psychiatric disorders including severe aggression/anti-social behavior [186], [187]. VPA often used as a mood stabilizer/in epilepsy, exerts its effect is through regulating the brain levels of serotonin. VPA activates monoamine oxidase (MAO) A catalytic activity via Akt/FoxO1 signaling pathway that degrades a number of monoamine neurotransmitters, including serotonin [188]. Therefore, VPA also acts as an anti-cancer agent through the serotonin pathway.

## II. CONCLUSION

Apart from the available chemotherapy, radiotherapy and surgery, the following seven steps have been suggested in this review study, if taken care of, may perhaps help to reduce the morbidity and mortality of a dreaded disease, cancer.

1. Estimation of Platelet 5HT level which is a Stress marker. Treatment of Mental Stress with medicine [189].
2. ATF3 dampening in the host may be tried as a potential therapeutic approach.
3. a) PLA to be measured routinely in cancer patients.
- b) NF- $\kappa$ B inhibitors may be used selectively as anti-cancer therapy e.g. Disulfiram, olmesartan etc.
- c) Selective blockage of LPA<sub>1</sub> and LPA<sub>2</sub> may be tried.
4. i) Drugs targeting the serotonin-5-HT<sub>2B</sub>R-FOXO3a pathway and normalising 5HT level to be used.
- ii) Agents that modulate the metabolism of serotonin may be useful for cancer treatment.
- iii) 5-HT1A, 5-HT1B, 5HT<sub>2A</sub>, 5HT<sub>2B</sub>, 5HT<sub>4</sub> and 5HT<sub>6</sub> effectively blocked the growth promoting effects of serotonin and selective blocking of these receptors may act as potential antineoplastic agents.
5. Inhibiting SKP2 may be tried for the treatment of different types of tumors.
6. Melatonin level to be measured routinely in cancer patients. Melatonin supplementation, due to its oncostatic properties, will immensely add on to the improvements with conventional cancer treatment.
7. Chronic VPA administration may result in profound decreases in proliferation in cancer. Different HDACI have been approved as anti-cancer agents.

## REFERENCES

- [1] Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer*. 2004; 4(8):579–591.
- [2] Chida Y, Hamer M, Wardle J, Steptoe A. Do stress-related psychosocial factors contribute to cancer incidence and survival? *Nat ClinPractOncol*. 2008; 5(8):466–475.
- [3] Pandey V, Vijayakumar MV, Ajay AK, Malvi P, Bhat MK. Diet-induced obesity increases melanoma progression: involvement of Cav-1 and FASN. *Int J Cancer*. 2012;130(3):497–508.
- [4] Sloan EK, et al. The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res*. 2010; 70(18):7042–7052.
- [5] Hai T, Hartman MG (2001) The molecular biology and nomenclature of the ATF/CREB family of transcription factors: ATF proteins and homeostasis. *Gene* 273:1–11.
- [6] Montminy M (1997) Transcriptional regulation by cyclic AMP. *Annu Rev Biochem* 66: 807–822.
- [7] Hai T, Wolfgang CD, Marsee DK, Allen AE, Sivaprasad U (1999) ATF3 and stress responses. *Gene Expr* 7:321–335.
- [8] Hai T, Wolford CC, Chang YS (2010) ATF3, a hub of the cellular adaptive-response network, in the pathogenesis of diseases: Is modulation of inflammation a unifying component? *Gene Expr* 15:1–11.
- [9] Balkwill F, Mantovani A (2001) Inflammation and cancer: Back to Virchow? *Lancet* 357:539–545.
- [10] Coussens LM, Werb Z (2002) Inflammation and cancer. *Nature* 420:860–867.
- [11] Mantovani A, Allavena P, Sica A, Balkwill F (2008) Cancer-related inflammation. *Nature* 454:436–444.
- [12] Tan TT, Coussens LM (2007) Humoral immunity, inflammation and cancer. *Curr Opin Immunol* 19:209–216.
- [13] Wolford CC, et al. (2013) Transcription factor ATF3 links host adaptive response to breast cancer metastasis. *J Clin Invest* 123:2893–2906.
- [14] Oh YK, et al. (2008) Role of activating transcription factor 3 on TAp73

- stability and apoptosis in paclitaxel-treated cervical cancer cells. *Mol Cancer Res* 6:1232–1249.
- [15] St Germain C, et al. (2010) Cisplatin induces cytotoxicity through the mitogenactivated protein kinase pathways and activating transcription factor 3. *Neoplasia* 12:527–538.
  - [16] Park EJ, Kwon HK, Choi YM, Shin HJ, Choi S (2012) Doxorubicin induces cytotoxicity through upregulation of pERK-dependent ATF3. *PLoS One* 7:e44990.
  - [17] Yi Seok Chang et al. Stress-inducible gene Atf3 in the noncancer host cells contributes to chemotherapy-exacerbated breast cancer metastasis. *PNAS* 2017 114 (34) E7159-E7168
  - [18] Emmenegger U, Kerbel RS (2010) Cancer: Chemotherapy counteracted. *Nature* 468: 637–638.
  - [19] Gilbert LA, Hemann MT (2011) chemotherapeutic resistance: Surviving stressful situations. *Cancer Res* 71:5062–5066.
  - [20] Camacho A, Dimsdale JE. Platelets and psychiatry: lessons learned from old and new studies. *Psychosom Med.* 2000;62:326–336.
  - [21] Halaris A. Comorbidity between depression and cardiovascular disease. *IntAngible.* 2009; 28:92–99.
  - [22] Bruce EC, Musselman DL. Depression, alterations in platelet function, and ischemic heart disease. *Psychosom Med.* 2005;67 Suppl 1:S34–S36.
  - [23] Schins A, Honig A, Crijns H, Baur L, Hamulyák K. Increased coronary events in depressed cardiovascular patients: 5-HT<sub>2A</sub> receptor as missing link. *Psychosom Med.* 2003;65:729–737.
  - [24] Nemeroff CB, Musselman DL. Are platelets the link between depression and ischemic heart disease. *Am Heart J.* 2000;140:57–62.
  - [25] Jurk K, Kehrel BE. Platelets: physiology and biochemistry. *SeminThrombHemost.* 2005;31:381–392.
  - [26] El-Sayed MS. Exercise and training effects on platelets in health and disease. *Platelets.* 2002; 13:261–266.
  - [27] Michelson AD. Flow cytometry: a clinical test of platelet function. *Blood.* 1996;87:4925–4936.
  - [28] Von Känel R. Platelet hyperactivity in clinical depression and the beneficial effect of antidepressant drug treatment: how strong is the evidence. *ActaPsychiatr Scand.* 2004;110:163–177.
  - [29] Von Känel R, Dimsdale JE. Effects of sympathetic activation by adrenergic infusions on hemostasis in vivo. *Eur J Haematol.* 2000; 65:357–369.
  - [30] Li N. Platelet-lymphocyte cross-talk. *J Leukoc Biol.* 2008;83:1069–1078.
  - [31] Siegel-Axel DI, Gawaz M. Platelets and endothelial cells. *SeminThrombHemost.* 2007; 33:128–135.
  - [32] Michelson AD. Methods for the measurement of platelet function. *Am J Cardiol.* 2009; 103:20A–26A.
  - [33] Hamer M, Gibson EL, Vuononvirta R, Williams E, Steptoe A. Inflammatory and hemostatic responses to repeated mental stress: individual stability and habituation over time. *Brain Behav Immun.* 2006;20:456–459.
  - [34] Steptoe A, Magid K, Edwards S, Brydon L, Hong Y, Eruslimsky J. The influence of psychological stress and socioeconomic status on platelet activation in men. *Atherosclerosis.* 2003; 168:57–63.
  - [35] Verheul HM, Jorna AS, Hoekman K, Broxterman HJ, Gebbink MF, Pinedo HM. Vascular endothelial growth factor-stimulated endothelial cells promote adhesion and activation of platelets. *Blood* 2000;96:4216–4221.
  - [36] Kisucka J, Butterfield CE, Duda DG, et al. Platelets and platelet adhesion support angiogenesis while preventing excessive hemorrhage. *Proceedings of the National Academy of Sciences of the United States of America* 2006; 103:855–860.
  - [37] Feng W, Madajka M, Kerr BA, Mahabeshwar GH, Whiteheart SW, Byzova TV. A novel role for platelet secretion in angiogenesis: mediating bone marrow-derived cell mobilization and homing. *Blood* 2011; 117:3893–3902.
  - [38] Goerge T, Ho-Tin-Noe B, Carbo C, et al. Inflammation induces hemorrhage in thrombocytopenia. *Blood* 2008; 111:4958–4964.
  - [39] Ho-Tin-Noe B, Goerge T, Cifuni SM, Duerschmied D, Wagner DD. Platelet granule secretion continuously prevents intratumor hemorrhage. *Cancer research* 2008; 68:6851–6858.
  - [40] Brock TA, Dvorak HF, Senger DR. Tumor-secreted vascular permeability factor increases cytosolic Ca<sup>2+</sup> and von Willebrand factor release in human endothelial cells. *Am J Pathol* 1991; 138:213–21.
  - [41] Pearlstein E, Salk PL, Yogeewaran G, Karpatkin S. Correlation between spontaneous metastatic potential, platelet-aggregating activity of cell surface extracts, and cell surface sialylation in 10 metastatic variant derivatives of a rat renal sarcoma cell line. *Proceedings of the National Academy of Sciences of the United States of America* 1980; 77:4336–4339.
  - [42] Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *Proceedings of the National Academy of Sciences of the United States of America* 1968; 61:46–52, 48.
  - [43] Kimoto M, Ando K, Koike S, et al. Significance of platelets in an antimetastatic activity of bacterial lipopolysaccharide. *Clinical & experimental metastasis* 1993;11:285–292.
  - [44] Nieswandt B, Hafner M, Echtenacher B, Mannel DN. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. *Cancer research* 1999; 59:1295–1300.
  - [45] Kopp HG, Placke T, Salih HR. Platelet-derived transforming growth factor-beta down-regulates NKG2D thereby inhibiting natural killer cell antitumor reactivity. *Cancer research* 2009; 69:7775–7783.
  - [46] Skov Madsen P, Hokland P, Hokland M. Secretory products from thrombin-stimulated human platelets exert an inhibitory effect on NK cytotoxic activity. *Acta pathologica, microbiologica, et immunologica Scandinavica Section C, Immunology* 1986;94:193–200.
  - [47] Laubli H, Borsig L. Selectins promote tumor metastasis. *Seminars in cancer biology* 2010; 20:169–177. 92. Labelle M, Begum S, Hynes RO. Direct signaling between platelets and cancer cells induces an epithelial-mesenchymal-like transition and promotes metastasis. *Cancer cell* 2011; 20:576–590.
  - [48] Felding-Habermann B, Habermann R, Saldivar E, Ruggeri ZM. Role of beta3 integrins in melanoma cell adhesion to activated platelets under flow. *The Journal of biological chemistry* 1996; 271:5892–5900.
  - [49] Cedervall J, Olsson A-K. Platelet Regulation of Angiogenesis, Tumor Growth and Metastasis. In: Ran S, ed. *Tumor Angiogenesis*, 2012: 115–134.
  - [50] Labelle M, Begum S, Hynes RO. Direct signaling between platelets and cancer cells induces an epithelial-mesenchymal-like transition and promotes metastasis. *Cancer cell* 2011; 20:576–590.
  - [51] Escárcega RO, Fuentes-Alexandro S, García-Carrasco M, Gatica A, Zamora A (March 2007). "The transcription factor nuclear factor-kappa B and cancer". *Clinical Oncology.* 19 (2): 154–61.
  - [52] Hamdy NA (January 2008). "Denosumab: RANKL inhibition in the management of bone loss". *Drugs of Today.* 44 (1): 7–21.
  - [53] Boucharaba A, Serre CM, Grès S, et al. Platelet-derived lysophosphatidic acid supports the progression of osteolytic bone metastases in breast cancer. *J Clin Invest* 2004; 114:1714–25.
  - [54] Belloc C, Lu H, Soria C, et al. The effect of platelets on invasiveness and protease production of human mammary tumor cells. *Int J Cancer* 1995;60:413–7.
  - [55] Gerrard JM, Robinson P. Identification of the molecular species of lysophosphatidic acid produced when platelets are stimulated by thrombin. *BiochimBiophysActa* 1989; 1001:282–5.
  - [56] Eichholtz T, Jalink K, Fahrenfort I, et al. The bioactive phospholipid lysophosphatidic acid is released from activated platelets. *Biochem J* 1993; 291:677–80.
  - [57] Sutphen R, Xu Y, Wilbanks GD, et al. Lysophospholipids are potential biomarkers of ovarian cancer. *Cancer Epidemiol Biomarkers Prev* 2004;13:1185–91.
  - [58] Merchant TE, Kasimos JN, de Graaf PW, et al. Phospholipid profiles of human colon cancer using 31P magnetic resonance spectroscopy. *Int J Colorectal Dis* 1991;6:121–6.
  - [59] Mills GB, Moolenaar WH. The emerging role of lysophosphatidic acid in cancer. *Nat Rev Cancer* 2003;3:582–91.
  - [60] Leblanc R, Peyruchaud O. New insights into the autotaxin/LPA axis in cancer development and metastasis. *Exp Cell Res* 2015;333:183–9.
  - [61] Boucharaba A, Serre CM, Guglielmi J, et al. The type 1 lysophosphatidic acid receptor is a target for therapy in bone metastases. *Proc Natl Acad Sci U S A* 2006; 103:9643–8.
  - [62] David M, Ribeiro J, Descotes F, et al. Targeting lysophosphatidic acid receptor type 1 with Debio 0719 inhibits spontaneous metastasis dissemination of breast cancer cells independently of cell proliferation and angiogenesis. *Int J Oncol* 2012; 40:1133–41.
  - [63] Yu S, Murph MM, Lu Y, et al. Lysophosphatidic acid receptors determine tumorigenicity and aggressiveness of ovarian cancer cells. *J Natl Cancer Inst* 2008;100:1630–42.
  - [64] Fishman DA, Liu Y, Ellerbroek SM, et al. Lysophosphatidic acid promotes matrix metalloproteinase (MMP) activation and MMP-dependent invasion in ovarian cancer cells. *Cancer Res* 2001;61:3194–9.
  - [65] Jeong KJ, Park SY, Cho KH, et al. The Rho/ROCK pathway for lysophosphatidic acid-induced proteolytic enzyme expression and ovarian cancer cell invasion. *Oncogene* 2012; 31:4279–89.

- [66] Park SY, Jeong KJ, Panupinthu N, et al. Lysophosphatidic acid augments human hepatocellular carcinoma cell invasion through LPA1 receptor and MMP-9 expression. *Oncogene* 2011;30:1351-9.
- [67] Hope JM, Wang FQ, Whyte JS, et al. LPA receptor 2 mediates LPA-induced endometrial cancer invasion. *GynecolOncol*2009;112:215-23.
- [68] Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol*2001;17:463-516.
- [69] Deryugina EI, Quigley JP. Matrix metalloproteinases and tumor metastasis. *Cancer Metastasis Rev* 2006;25:9-34.
- [70] Ho-Tin-Noé B, Goerge T, Cifuni SM, et al. Platelet granule secretion continuously prevents intratumor hemorrhage. *Cancer Res* 2008;68:6851-8.
- [71] Hettiarachchi RJ, Smorenburg SM, Ginsberg J, et al. Do heparins do more than just treat thrombosis? The influence of heparins on cancer spread. *ThrombHaemost*1999;82:947-52.
- [72] Borly L, Wille-Jørgensen P, Rasmussen MS. Systematic review of thromboprophylaxis in colorectal surgery -- an update. *Colorectal Dis* 2005;7:122-7.
- [73] Smorenburg SM, Hettiarachchi RJ, Vink R, et al. The effects of unfractionated heparin on survival in patients with malignancy--a systematic review. *Thromb Haemost*1999; 82:1600-4.
- [74] Akl EA, van Doormaal FF, Barba M, et al. Parenteral anticoagulation may prolong the survival of patients with limited small cell lung cancer: a Cochrane systematic review. *J Exp Clin Cancer Res*2008; 27:4.
- [75] Nierodzic ML, Klepfish A, Karpatic S. Role of platelets, thrombin, integrin IIb-IIIa, fibronectin and von Willebrand factor on tumor adhesion in vitro and metastasis in vivo. *ThrombHaemost* 1995; 74:282-90.
- [76] Hiroshi Kawahara et al. Psychological stress increases serotonin release in the rat amygdala and prefrontal cortex assessed by in vivo micro dialysis. *Neuroscience Letters* 1993; Volume 162, Issues 1-2: 81-84.
- [77] Mitsuhiro Yoshioka et al. Effects of conditioned fear stress on 5-HT release in the rat prefrontal cortex. *Pharmacology Biochemistry and Behavior*; June-July 1995, Volume 51, Issues 2-3, Pages 515-519.
- [78] Francis Chaouloff et al. Serotonin and Stress. *Neuropsychopharmacology* (1999) 21, 28S-32S.
- [79] Chaouloff F. (1993): Physiopharmacological interactions between stress hormones and central serotonergic systems. *Brain Res Rev* 18: 1-32.
- [80] Chao Liang et al. Serotonin promotes the proliferation of serum-deprived hepatocellular carcinoma cells via upregulation of FOXO3a. *Molecular Cancer* 2013, 12:14.
- [81] Soll C et al. Serotonin promotes tumor growth in human hepatocellular cancer. *Hepatology*. 2010 Apr; 51(4):1244-54.
- [82] Siddiqui EJ et al. The role of serotonin (5-hydroxytryptamine1A and 1B) receptors in prostate cancer cell proliferation. *J Urol*. 2006 Oct;176(4 Pt 1):1648-53.
- [83] Dizéy N et al. Serotonin activates MAP kinase and PI3K/Akt signaling pathways in prostate cancer cell lines. *Urol Oncol*. 2011 Jul-Aug;29(4):436-45.
- [84] Gianfranco Alpini et al. Serotonin metabolism is dysregulated in cholangiocarcinoma, which has implications for tumor growth *Cancer Res*. 2008 Nov 15; 68(22): 9184-9193.
- [85] Antonio Nocito et al. Serotonin Regulates Macrophage-Mediated Angiogenesis in a Mouse Model of Colon Cancer Allografts. *Cancer Res* July 1, 2008 68; 5152-8.
- [86] Houghton AM, Grisolan JL, Baumann ML, et al. Macrophage elastase (matrix metalloproteinase-12) suppresses growth of lung metastases. *Cancer Res* 2006; 66: 6149-55.
- [87] Shapiro SD. Diverse roles of macrophage matrix metalloproteinases in tissue destruction and tumor growth. *ThrombHaemost* 1999; 82: 846-9.
- [88] Cattaneo MG, Codignola A, Vicentini LM, Clementi F, Sher E. Nicotine stimulates a serotonergic autocrine loop in human small-cell lung carcinoma. *Cancer Res* 1993; 53: 5566-8.
- [89] Pratesi G, Cervi S, Balsari A, Bondiolotti G, Vicentini LM. Effect of serotonin and nicotine on the growth of a human small cell lung cancer xenograft. *Anticancer Res* 1996; 16: 3615-9.
- [90] Pelagio-Flores R. et al. Serotonin, a tryptophan-derived signal conserved in plants and animals, regulates root system architecture probably acting as a natural auxin inhibitor in *Arabidopsis thaliana*. *Plant Cell Physiol*. 2011 Mar;52(3):490-508.
- [91] Yu-Shan Chen et al. Ubiquitin at the crossroad of cell death and survival. *Chin J Cancer*. 2013 Dec; 32(12): 640-647.
- [92] Jerry Vriend et al. Breast cancer cells: Modulation by melatonin and the ubiquitin-proteasome system – A review. *Molecular and Cellular Endocrinology*. Volume 417, 5 December 2015, Pages 1-9.
- [93] Wäsch R, Engelbert D. Anaphase-promoting complex-dependent proteolysis of cell cycle regulators and genomic instability of cancer cells. *Oncogene*. 2005;24:1-10.
- [94] Hoeller D et al.. Ubiquitin and ubiquitin-like proteins in cancer pathogenesis. *Nat Rev Cancer*. 2006;6:776-788.
- [95] Adams J. The development of proteasome inhibitors as anticancer drugs. *Cancer cell*.2004;5:417-421.
- [96] Wang Z, Liu P et al. Wei W. Roles of F-box proteins in cancer. *Nat Rev Cancer* 2014; 14:233-247.
- [97] Carrano AC, Eytan E, Hershko A, Pagano M. SKP2 is required for ubiquitin-mediated degradation of the CDK inhibitor p27. *Nat Cell Biol* 1999; 1:193-199.
- [98] Sutterluty H, Chatelain E, Marti A, et al. p45SKP2 promotes p27Kip1 degradation and induces S phase in quiescent cells. *Nat Cell Biol* 1999; 1:207-214.
- [99] Yu ZK, Gervais JL, Zhang H. Human CUL-1 associates with the SKP1/SKP2 complex and regulates p21(CIP1/WAF1) and cyclin D proteins. *ProcNatAcadSci USA* 1998; 95:11324-11329.
- [100] Chan CH, Li CF, Yang WL, et al. The Skp2-SCF E3 ligase regulates Aktubiquitination, glycolysis, hereceptin sensitivity, and tumorigenesis. *Cell* 2012; 149:1098-1111.
- [101] Radke S, Pirkmaier A, Germain D. Differential expression of the F-box proteins Skp2 and Skp2B in breast cancer. *Oncogene* 2005; 24:3448-3458.
- [102] Li J-Q, Wu F, Masaki T, et al. Correlation of Skp2 with carcinogenesis, invasion, metastasis, and prognosis in colorectal tumors. *Int J Oncol* 2004; 25:87-95.
- [103] Seki R et al. Prognostic significance of S-phase kinase-associated protein 2 and p27kip1 in patients with diffuse large B-cell lymphoma: effects of rituximab. *Ann Oncol* 2010; 21:833-841.
- [104] Gstaiger M, Jordan R, Lim M, et al. Skp2 is oncogenic and overexpressed in human cancers. *ProcNatAcadSci USA* 2001; 98:5043-5048.
- [105] Lu M, Ma J, Xue W, et al. The expression and prognosis of FOXO3a and Skp2 in human hepatocellular carcinoma. *PatholOncol Res* 2009; 15:679-687.
- [106] Ester K et al. The phytohormone auxin induces G1 cell-cycle arrest of human tumor cells. *Planta Med*. 2009 Oct;75(13):1423-6.
- [107] Dominguez-Rodriguez A, Breu-Gonzalez P (2011) Melatonin: still a forgotten antioxidant. *International Journal of Cardiology* 149: 382.
- [108] Korkmaz A, Reiter RJ, Topal T, Manchester LC, Oter S, et al. (2009) Melatonin: an established antioxidant worthy of use in clinical trials. *Molecular Medicine* 15: 43-50.
- [109] Galano A, Tan DX, Reiter RJ (2011) Melatonin as a natural ally against oxidative stress: a physicochemical examination. *Journal of Pineal Research* 51: 1-16.
- [110] Bonnefont-Rousselot D, Collin F, Jore D, Gardès-Albert M (2011) Reaction mechanism of melatonin oxidation by reactive oxygen species in vitro. *Journal of Pineal Research* 50: 328-335.
- [111] Reiter RJ, Tan DX, Poeggeler B, Menendez-Pelaez, Chen L, et al. (1994) Melatonin as a free radical scavenger: implications for aging and age-related diseases. *Annals of The New York Academy of Sciences* 719: 1-12.
- [112] Tan DX, Chen LD, Poeggeler B, Manchester LC, Reiter RJ, et al. (1993) Melatonin: a potent endogenous hydroxyl radical scavenger. *Endocrine Journal* 1: 57-60.
- [113] Hill SM, Blask DE, Xiang S, Yuan L, Mao L, et al. (2011) Melatonin and associated signaling pathways that control normal breast epithelium and breast cancer. *Journal of Mammary Gland Biology and Neoplasia* 16: 235-245.
- [114] Messina G, Lissoni P, Marchiori P, Bartolacelli E, Brivio F, et al. (2010) Enhancement of the efficacy of cancer chemotherapy by the pineal hormone melatonin and its relation with the psychospiritual status of cancer patients. *Journal of Research in Medical Sciences* 15: 225-228.
- [115] Padillo FJ, Ruiz-Rabelo JF, Cruz A, Perea MD, Tasset I, et al. (2010) Melatonin and celecoxib improve the outcomes in hamsters with experimental pancreatic cancer. *Journal of Pineal Research* 49: 264-270.
- [116] Grant SG, Melan MA, Latimer JJ, Witt-Enderby PA (2009) Melatonin and breast cancer: cellular mechanisms, clinical studies and future perspectives. *Expert Reviews in Molecular Medicine* 11: e5.
- [117] Srinivasan V, Spence DW, Pandi-Perumal SR, Trakht I, Cardinali DP, et al. (2008) Therapeutic actions of melatonin in cancer: possible mechanisms. *Integrative Cancer Therapies* 7: 189-203.
- [118] Garcia-Navarro A, Gonzalez-Puga C, Escames G, López LC, López A, et al. (2007) Cellular mechanisms involved in the melatonin inhibition of

- HT-29 human colon cancer cell proliferation in culture. *Journal of Pineal Research* 43: 195–205.
- [119] Jung-Hynes B, Reiter RJ, Ahmad N (2010) Sirtuins, melatonin and circadian rhythms: building a bridge between aging and cancer. *Journal of Pineal Research* 48: 9–19.
- [120] Gonzalez A, Del Castillo-Vaquero A, Miro-Moran A, Tapia JA, Salido GM, et al. (2011) Melatonin reduces pancreatic tumor cell viability by altering mitochondrial physiology. *Journal of Pineal Research* 50: 250–260.
- [121] Um HJ, Park JW, Kwon TK (2011) Melatonin sensitizes Caki renal cancer cells to kahweol-induced apoptosis through CHOP mediated up-regulation of PUMA. *Journal of Pineal Research* 50: 359–366.
- [122] Mao L, Yuan L, Slakey LM, Jones FE, Burow ME, et al. (2010) Inhibition of breast cancer cell invasion by melatonin is mediated through regulation of the p38 mitogen-activated protein kinase signaling pathway. *Breast Cancer Research* 12: R107.
- [123] Proietti S, Cucina A, D'Anselmi F, Dinicola S, Pasqualato A, et al. (2011) Melatonin and vitamin D3 synergistically down-regulate Akt and MDM2 leading to TGFbeta-1-dependent growth inhibition of breast cancer cells. *Journal of Pineal Research* 50: 150–158.
- [124] Martínez-Campa CM, Alonso-González C, Mediavilla D, Cos S, González A, et al. (2008) Melatonin down-regulates hTERT expression induced by either natural estrogens (17beta-estradiol) or metalloestrogens (cadmium) in MCF-7 human breast cancer cells. *Cancer Letters* 268: 272–277.
- [125] Dai M, Cui P, Yu M, Han J, Li H, et al. (2008) Melatonin modulates the expression of VEGF and HIF-1 alpha induced by CoCl<sub>2</sub> in cultured cancer cells. *Journal of Pineal Research* 44: 121–126.
- [126] Plaimée P, Weerapreeyakul N, Barusruks S, Johns NP. Melatonin potentiates cisplatin-induced apoptosis and cell cycle arrest in human lung adenocarcinoma cells. *Cell Prolif*. 2015;48:67–77.
- [127] Fan C, Pan Y, Yang Y, Di S, Jiang S, Ma Z, Li T, Zhang Z, Li W, Li X, Reiter RJ, Yan X. HDAC1 inhibition by melatonin leads to suppression of lung adenocarcinoma cells via induction of oxidative stress and activation of apoptotic pathways. *J Pineal Res*. 2015;59:321–333.
- [128] Plaimée P, Weerapreeyakul N, Thumanu K, Tanthanuch W, Barusruks S. Melatonin induces apoptosis through biomolecular changes in SK-LU-1 human lung adenocarcinoma cells. *Cell Prolif*. 2014;47:564–577.
- [129] Alvarez-Garcia V, Gonzalez A, Alonso-Gonzalez C, Martinez-Campa C, Cos S. Regulation of vascular endothelial growth factor by melatonin in human breast cancer cells. *J Pineal Res*. 2013;54:373–380.
- [130] Borin TF, Arbab AS, Gelaleti GB, Ferreira LC, Moschetta MG, Jardim-Perassi BV, Iskander A, Varma NR, Shankar A, Coimbra VB, Fabri VA, de Oliveira JG, Zuccari DA. Melatonin decreases breast cancer metastasis by modulating Rho-associated kinase protein-1 expression. *J Pineal Res*. 2016;60:3–15.
- [131] Woo SM, Min KJ, Kwon TK. Melatonin-mediated Bim up-regulation and cyclooxygenase-2 (COX-2) down-regulation enhances tunicamycin-induced apoptosis in MDA-MB-231 cells. *J Pineal Res*. 2015;58:310–320.
- [132] Alonso-Gonzalez C, Gonzalez A, Martinez-Campa C, Gomez-Arozamena J, Cos S. Melatonin sensitizes human breast cancer cells to ionizing radiation by downregulating proteins involved in double-strand DNA break repair. *J Pineal Res*. 2015;58:189–197.
- [133] Proietti S, Cucina A, Dobrowolny G, D'Anselmi F, Dinicola S, Masiello MG, Pasqualato A, Palombo A, Morini V, Reiter RJ, Bizzarri M. Melatonin down-regulates MDM2 gene expression and enhances p53 acetylation in MCF-7 cells. *J Pineal Res*. 2014;57:120–129.
- [134] Hevia D, Gonzalez-Menendez P, Quiros-Gonzalez I, Miar A, Rodriguez-Garcia A, Tan DX, Reiter RJ, Mayo JC, Sainz RM. Melatonin uptake through glucose transporters: a new target for melatonin inhibition of cancer. *J Pineal Res*. 2015;58:234–250.
- [135] Paroni R, Terraneo L, Bonomini F, Finati E, Virgili E, Bianciardi P, Favero G, Fraschini F, Reiter RJ, Rezzani R, Samaja M. Antitumour activity of melatonin in a mouse model of human prostate cancer: relationship with hypoxia signalling. *J Pineal Res*. 2014;57:43–52.
- [136] Shiu SY, Leung WY, Tam CW, Liu VW, Yao KM. Melatonin MT1 receptor-induced transcriptional up-regulation of p27(Kip1) in prostate cancer antiproliferation is mediated via inhibition of constitutively active nuclear factor kappa B (NF-kappaB): potential implications on prostate cancer chemoprevention and therapy. *J Pineal Res*. 2013;54:69–79.
- [137] Joo SS, Yoo YM. Melatonin induces apoptotic death in LNCaP cells via p38 and JNK pathways: therapeutic implications for prostate cancer. *J Pineal Res*. 2009;47:8–14.
- [138] Ordonez R, Fernandez A, Prieto-Dominguez N, Martinez L, Garcia-Ruiz C, Fernandez-Checa JC, Mauriz JL, Gonzalez-Gallego J. Ceramide metabolism regulates autophagy and apoptotic cell death induced by melatonin in liver cancer cells. *J Pineal Res*. 2015;59:178–189.
- [139] Ordonez R, Carbajo-Pescador S, Prieto-Dominguez N, Garcia-Palomo A, Gonzalez-Gallego J, Mauriz JL. Inhibition of matrix metalloproteinase-9 and nuclear factor kappa B contribute to melatonin prevention of motility and invasiveness in HepG2 liver cancer cells. *J Pineal Res*. 2014;56:20–30.
- [140] Leon J, Casado J, Jimenez Ruiz SM, Zurita MS, Gonzalez-Puga C, Rejon JD, Gila A, Munoz de Rueda P, Pavon EJ, Reiter RJ, Ruiz-Extremera A, Salmeron J. Melatonin reduces endothelin-1 expression and secretion in colon cancer cells through the inactivation of FoxO-1 and NF-kappabeta. *J Pineal Res*. 2014;56:415–426.
- [141] Hong Y, Won J, Lee Y, Lee S, Park K, Chang KT, Hong Y. Melatonin treatment induces interplay of apoptosis, autophagy, and senescence in human colorectal cancer cells. *J Pineal Res*. 2014;56:264–274.
- [142] Zhou Q, Gui S, Zhou Q, Wang Y. Melatonin inhibits the migration of human lung adenocarcinoma A549 cell lines involving JNK/MAPK pathway. *PLoS One*. 2014;9:e101132.
- [143] Plaimée P, Khamphio M, Weerapreeyakul N, Barusruks S, Johns NP. Immunomodulatory effect of melatonin in SK-LU-1 human lung adenocarcinoma cells co-cultured with peripheral blood mononuclear cells. *Cell Prolif*. 2014;47:406–415.
- [144] Lissoni P, Rovelli F, Malugani F, Bucovec R, Conti A, Maestroni GJ. Anti-angiogenic activity of melatonin in advanced cancer patients. *Neuro Endocrinol Lett*. 2001;22:45–47.
- [145] Mocchegiani E, Perissin L, Santarelli L, Tibaldi A, Zorzet S, Rapozzi V, Giacconi R, Bulian D, Giraldi T. Melatonin administration in tumor-bearing mice (intact and pinealectomized) in relation to stress zinc thymulin and IL-2. *Int J Immunopharmacol*. 1999;21:27–46.
- [146] Lissoni P, Chillelli M, Villa S, Cerizza L, Tancini G. Five years survival in metastatic non-small cell lung cancer patients treated with chemotherapy alone or chemotherapy and melatonin: a randomized trial. *J Pineal Res*. 2003;35:12–15.
- [147] Lissoni P. Biochemotherapy with standard chemotherapies plus the pineal hormone melatonin in the treatment of advanced solid neoplasms. *Pathol Biol (Paris)* 2007;55:201–204.
- [148] Cos S, Fernández R, Guezmes A, Sánchez-Barceló EJ (1998) Influence of melatonin on invasive and metastatic properties of MCF-7 human breast cancer cells. *Cancer research* 58: 4383–4390.
- [149] Ortiz-Lopez L, Morales-Mulia S, Ramirez-Rodriguez G, Benítez-King G (2009) ROCK-regulated cytoskeletal dynamics participate in the inhibitory effect of melatonin on cancer cell migration. *Journal of Pineal Research* 46: 15–21.
- [150] Ramirez-Rodriguez G, Ortiz-Lopez L, Benitez-King G (2007) Melatonin increases stress fibers and focal adhesions in MDCK cells: participation of Rho-associated kinase and protein kinase C. *Journal of Pineal Research*. 42: 180–190.
- [151] Bellon A, Ortiz-Lopez L, Ramirez-Rodriguez G, Antón-Tay F, Benitez-King G (2007) Melatonin induces neuritegenesis at early stages in N1E-115 cells through actin rearrangements via activation of protein kinase C and Rho-associated kinase. *Journal of Pineal Research* 42: 214–221.
- [152] Q Zhou et al. Melatonin Inhibits the Migration of Human Lung Adenocarcinoma A549 Cell Lines Involving JNK/MAPK Pathway. *journals.plos.org*. Jul 3, 2014.
- [153] Pariente R, Pariente JA, Rodriguez AB, Espino J. Melatonin sensitizes human cervical cancer HeLa cells to cisplatin-induced cytotoxicity and apoptosis: effects on oxidative stress and DNA fragmentation. *J Pineal Res*. 2016;60:55–64.
- [154] Leja-Szpak A, Jaworek J, Pierzchalski P, Reiter RJ. Melatonin induces pro-apoptotic signaling pathway in human pancreatic carcinoma cells (PANC-1) *J Pineal Res*. 2010;49:248–255.
- [155] Park EJ, Woo SM, Min KJ, Kwon TK. Transcriptional and post-translational regulation of Bim controls apoptosis in melatonin-treated human renal cancer Caki cells. *J Pineal Res*. 2014;56:97–106.
- [156] Garcia-Santos G, Antolin I, Herrera F, Martin V, Rodriguez-Blanco J, del Pilar Carrera M, Rodriguez C. Melatonin induces apoptosis in human neuroblastoma cancer cells. *J Pineal Res*. 2006;41:130–135.
- [157] Xin Z, Jiang S, Jiang P, Yan X, Fan C, Di S, Wu G, Yang Y, Reiter RJ, Ji G. Melatonin as a treatment for gastrointestinal cancer: a review. *J Pineal Res*. 2015;58:375–387.
- [158] Bizzarri M, Proietti S, Cucina A, Reiter RJ. Molecular mechanisms of the pro-apoptotic actions of melatonin in cancer: a review. *Expert Opin Ther Targets*. 2013;17:1483–1496.
- [159] Fernandez A, Ordonez R, Reiter RJ, Gonzalez-Gallego J, Mauriz JL.



- Melatonin and endoplasmic reticulum stress: relation to autophagy and apoptosis. *J Pineal Res.* 2015;59:292–307.
- [160] Vriend J, Reiter RJ. Melatonin as a proteasome inhibitor. Is there any clinical evidence? *Life Sci.* 2014;115(12):8–14.
- [161] Jaworek J, Leja-Szpak A. Melatonin influences pancreatic cancerogenesis. *Histol Histopathol.* 2014;29:423–431.
- [162] Rodriguez C, Martin V, Herrera F, Garcia-Santos G, Rodriguez-Blanco J, Casado-Zapico S, Sanchez-Sanchez AM, Suarez S, Puente-Moncada N, Anitua MJ, Antolin I. Mechanisms involved in the pro-apoptotic effect of melatonin in cancer cells. *Int J Mol Sci.* 2013;14:6597–6613.
- [163] Proietti S, Cucina A, Reiter RJ, Bizzarri M. Molecular mechanisms of melatonin's inhibitory actions on breast cancers. *Cell Mol Life Sci.* 2013;70:2139–2157.
- [164] Sanchez-Hidalgo M, Guerrero JM, Villegas I, Packham G, de la Lastra CA. Melatonin a natural programmed cell death inducer in cancer. *Curr Med Chem.* 2012;19:3805–3821.
- [165] Lanoix D, Lacasse AA, Reiter RJ, Vaillancourt C. Melatonin: the smart killer: the human trophoblast as a model. *Mol Cell Endocrinol.* 2012;348:1–11.
- [166] Mediavilla MD, Sanchez-Barcelo EJ, Tan DX, Manchester L, Reiter RJ. Basic mechanisms involved in the anti-cancer effects of melatonin. *Curr Med Chem.* 2010;17:4462–4481.
- [167] Sainz RM, Mayo JC, Rodriguez C, Tan DX, Lopez-Burillo S, Reiter RJ. Melatonin and cell death: differential actions on apoptosis in normal and cancer cells. *Cell Mol Life Sci.* 2003;60:1407–1426.
- [168] RJ Reiter et al. Melatonin, a Full Service Anti-Cancer Agent: Inhibition of Initiation, Progression and Metastasis. *International Journal of Molecular Sciences.* Apr 17, 2017.
- [169] Daniel Cardinal et al. Melatonin-Induced Oncostasis, Mechanisms and Clinical Relevance. *Journal of Integrative Oncology.* February 19, 2016
- [170] Tomas Eckschlag et al. Histone Deacetylase Inhibitors as Anticancer Drugs. *Int. J. Mol. Sci.* 2017, 18, 1414.
- [171] Chen, C. L., Sung, J et al. Valproic acid inhibits invasiveness in bladder cancer but not in prostate cancer cells. *J. Pharmacol. Exp. Ther.* 2006, 319, 533–542.
- [172] Vrana, J. A et al. Induction of apoptosis in U937 human leukemia cells by suberoylanilide hydroxamic acid (SAHA) proceeds through pathways that are regulated by Bcl-2/Bcl-XL, c-Jun, and p21CIP1, but independent of p53. *Oncogene* 1999, 18, 7016–7025.
- [173] Richon, V. M. et al. Histone deacetylase inhibitor selectively induces p21WAF1 expression and gene-associated histone acetylation. *Proc. Natl. Acad. Sci. USA* 2000, 97, 10014–10019.
- [174] Sander, V. et al. P21-dependent G1 arrest with downregulation of cyclin D1 and upregulation of cyclin E by the histone deacetylase inhibitor FR901228. *Br. J. Cancer* 2000, 83, 817–825.
- [175] Kim, H. J.; Bae, S.C. Histone deacetylase inhibitors: Molecular mechanisms of action and clinical trials as anti-cancer drugs. *Am. J. Transl. Res.* 2011, 3, 166–179.
- [176] Minucci, S.; Pelicci, P.G. Histone deacetylase inhibitors and the promise of epigenetic (and more) treatments for cancer. *Nat. Rev. Cancer* 2006, 6, 38–51.
- [177] Miller, C. P. et al. Therapeutic strategies to enhance the anticancer efficacy of histone deacetylase inhibitors. *J. Biomed. Biotechnol.* 2011, 2011, 514261.
- [178] Chiao, M. T., et al. Suberoylanilide hydroxamic acid (SAHA) causes tumor growth slowdown and triggers autophagy in glioblastoma stem cells. *Autophagy* 2013, 9, 1509–1526.
- [179] Gottlicher M, Minucci S, Zhu P, et al. Valproic acid defines a novel class of HDAC inhibitors inducing differentiation of transformed cells. *EMBO J* 2001; 20: 6969–78.
- [180] Salvador, M. A.; et al. The histone deacetylase inhibitor abexinostat induces Cancer stem cells differentiation in breast Cancer with low Xist expression. *Clin. Cancer Res.* 2013, 19, 6520–6531.
- [181] Yuan, P.X.; et al. The Mood Stabilizer Valproic Acid Activates Mitogen-activated Protein Kinases and Promotes Neurite Growth. *J. Biol. Chem.* 2001, 276, 31674–31683.
- [182] Cinatl, J.; et al. Induction of differentiation and suppression of malignant phenotype of human neuroblastoma BE(2)-C cells by valproic acid: Enhancement by combination with interferon- $\alpha$ . *Int. J. Oncol.* 2002, 20, 97–106.
- [183] Kroesen, M.; et al. HDAC inhibitors and immunotherapy; a double edged sword? *Oncotarget* 2014, 5, 6558–6572
- [184] Vivek Venkataramani et al. Histone Deacetylase Inhibitor Valproic Acid Inhibits Cancer Cell Proliferation via Down-regulation of the Alzheimer Amyloid Precursor Protein. *The Journal of Biological Chemistry.* April 2, 2010;285, 10678–10689.
- [185] Marks PA, Richon VM, Miller T, Kelly WK. Histone deacetylase inhibitors. *Adv Cancer Res* 2004; 91: 137–68.
- [186] Shih JC, Chen K, and Ridd MJ. Monoamine oxidase: from genes to behavior. *Annu Rev Neurosci.* 1999;22:197–217.
- [187] Bortolato M, Chen K, and Shih JC. Monoamine oxidase inactivation: from pathophysiology to therapeutics. *Adv Drug Deliv Rev.* 2008; 60:1527–1533.
- [188] Jason Boyang Wu and Jean C. Shih. Valproic Acid Induces Monoamine Oxidase A via Akt/Forkhead Box O1 Activation. *Mol Pharmacol* 80:714–723, 2011.
- [189] Devasis Ghosh. A Novel Method to combat stress by modulating Platelet serotonin levels by medicine. *Stress Management Professional* .2015;vol 3 (1);72-77.