# Effect of Drought Stress and Selenium Spraying on Superoxide Dismotase Activity of Winter Rapeseed (*Brassica napus* L.) Cultivars

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Abstract—In the other to Study of drought stress and Selenium spraying effect on superoxide dismotase (SOD) activity of rapeseed (Brassica napus L.) cultivars in Shahr-e-Rey region, an experiment carried out in Split factorial design in the basis of randomized complete blocks with 4 replications in 2006. Irrigation in two levels: Normal irrigation and irrigation with drought stress when the soil electrical conductivity reached to 60 as main factor and rapeseed cultivars in 3 levels Zarfam, Okapi, Opera and selenium spraying at the beginning of flowering stage in 3 levels: 0, 16 and 21 g/ha as sub factor

The results showed that the simple and interaction effect of irrigation, selenium and cultivars on SOD activity had significant difference. In this case Zarfam cultivar with 2010 u.mg<sup>-1</sup> protein and Opera with 1454 u.mg<sup>-1</sup> protein produced maximum and minimum amounts of SOD activitiy. Interaction effect of irrigation and variety showed that, normal irrigation in Opera with 1115 u.mg<sup>-1</sup> protein and drought stress in Zarfam with 2784 u.mg<sup>-1</sup> protein conducted to and minimum and maximum amounts of SOD activity.

Interaction effect of irrigation, cultivar and selenium on SOD indicated that drought stress condition and 21 gr/ha selenium spraying in Zarfam variety with 3146 u.mg<sup>-1</sup> protein gained to highest activities of SOD.

**Keywords**—Drought stress, Rapeseed, Selenium, Superoxide dismutase.

# I. INTRODUCTION

THE AOS such as  $O_2$ ,  $H_2O_2$  and OH radicals, can directly attack membrane lipids, inactive metabolic enzymes and damage the nucleic acids leading to cell death [12]. Being toxic for cells, AOS are efficiently eliminated by nonenzymatic ( $\alpha$ -tocopherol,  $\beta$  -carotene, phenolic

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compounds, ascorbate, glutathione) and enzymatic antioxidants [10]-[17]. The enzymatic antioxidant system is one of the protective mechanisms including superoxide dismutase (SOD: EC 1.15.1.1), which can be found in various cell compartments and it catalyses the isproportion of two  $O_2$  radicals to  $H_2O_2$  and  $O_2$  [5]-[14].

In the conditions of drought stress and because of that the aperture are close for preventing from the transpiration it would have concluded the reduction in the  $CO_2$ 's absorption and would have the less producing the amount of dry substance and it would use the food elements with the less efficiency. This counter effect for the amount of humidity and nutriment elements are bilateral[1]-[9].

Plants have developed the scavenging mechanism of ROS categorized as enzymatic and non-nzymatic[2]-[16]. When ROS increases, chain reactions start, in which Superoxide Dismutase (SOD) catalyzes the dismutation of  $\rm O_2^-$  radicals to molecular  $\rm O_2$  and  $\rm H_2O_2$ . The  $\rm H_2O_2$  is then detoxified in the ascorbate-glutathione cycle [12].

Selenium is an essential trace element for animals and humans [20] but its role in plants is still unclear [4]. Most cereal crops and fodder plant are relatively weakly able to absorb selenium, even when grown on soils with higher selenium content. Selenium is chemically similar to sulphur, this may cause a non-specific replacement of S by selenium in proteins and other sulphur compounds [11]. There are indications that it can also play a positive biological role in higher plants [3]. Selenium could increase the tolerance of plants to UV-radiation induced oxidative stress, delay senescence, and promote the growth of ageing seedlings [22]. The Results showed that plant growth promoted by selenium due to the increased starch accumulation chloroplasts[13]. Recently it has been shown that selenium can regulate the water status of plants under conditions of water deficiency and thereby performs its protective effect [7].

Se also decreased the activity of superoxide dismutase (SOD) and lowered the amount of tocopherols in some cases. It was suggested that while Se promotes  $H_2O_2$  scavenging, by increasing GPX activity, it also enhances spontaneous disproportion of superoxide radicals to  $H_2O_2$  and thus decreases the need for high SOD activity [22]. Se can alleviate oxidative stress in chloroplasts. The responses of potato to Se supplementation were investigated by monitoring chlorophyll

fluorescence and the transcription of antioxidative enzymes [15].

### II. MATERIALS AND METHODS

In the other to Study of drought stress and Selenium spraying on CAT and SOD activities of rapeseed (*Brassica napus* L.) cultivars in Shahr-e-Rey region, an experiment carried out in Split factorial design in the basis of randomized complete blocks with 4 replications in 2006. Irrigation in two levels: Normal irrigation and irrigation with drought stress when the soil electrical conductivity reached to 60 as main factor and rapeseed cultivars in 3 levels Zarfam, Okapi, Opera and selenium spraying at the beginning of flowering stage in 3 levels: 0, 16 and 21 g/ha as sub factor.

Seeds planted on 6 rows in each plots, the rows distance was 25 cm and the plant distance on each row was 4.5 cm, it had used 150 kg/ha pure nitrogen fertilizer and 60 kg/ha phosphorus and potassium, for maximum exact, beginning and end of each plots closed, with regarding area of each plots, irrigation intervals and water amounts used in each irrigation, volume of water for each plot determined.

In this research for uniform selenium spraying of each plot at first sprayer machine calibrated with water and sodium selenit treatment (0, 16 and 21 g/ha) sprayed by sodium selenit (Na203Se.5H20) form in early flowering stage.

First and last rows of each plot considered as edge effect and sampling for SOD enzyme activity did from 4 central rows (the area 4 m2 of each plot). SOD activity was assayed by measuring the ability of the enzyme extract to inhibit the photochemical reduction of NBT glass test tubes containing the mixture were illuminated with a fluorescent lamp (120 W); identical tubes that were not illuminated served as blanks. After illumination for 15 min, the absorbance was measured at 560 nm. One unit of SOD was defined as the amount of enzyme activity that was able to inhibit by 50% the photo reduction of NBT to blue Formosan. The SOD activity of the extract was expressed as SOD units per milligram of PROT. Peroxides activity was determined by the oxidation of guaiacol in the presence of H<sub>2</sub>O<sub>2</sub>. The increase in absorbance was recorded at 470 nm (Hernandez et al., 2000). The reaction mixture contained 100 μL crude enzyme, 500 μL H<sub>2</sub>O<sub>2</sub> 5 mM, 500 μL guaiacol 28 mM and 1900 μL potassium phosphate buffer 60 mM (pH 6.1). POX activity of the extract was expressed as POX units per mg [18].

## III. RESULTS AND DISCUSSION

Simple effect of irrigation, cultivar and foliar application of Selenium on SOD activity had significant difference at %1 level of probability (Table I). In this case drought stress condition with 2289.9 u.mg<sup>-1</sup>protein, Zarfam cultivar with 2010 u.mg<sup>-1</sup>protein and 21 g/ha Se application with 1973.0 u.mg<sup>-1</sup>protein showed maximum amounts of SOD activity (Table II).

TABLE I
ANALYSIS OF VARIANCE FOR SOD ACTIVITY

	M.S		
FACTOR	Degree of freedo m	SOD	
Replication	3	80429.389 <sup>ns</sup>	
Irrigation	1	18974853.389**	
Error	3	9155.759	
Cultivar	2	199694.347**	
Selenium	2	1237121.347**	
cultivar * Irrigation	9	1267009.847**	
Selenium * Irrigation	9	459804.514**	
Selenium * Cultivar	4	9243.701*	
Selenium *Cultivar *Irrigation	4	1952.785 *	
Error	48	5754.022	
Total	71	-	
C.V(%)	-	7.27	

ns, \* and \*\*:Nonsignificant and significant at %5 and %1 level of probability respectively

TABLE II MEAN COMPARISON OF IRRIGATION, VARIETY AND SELENIUM EFFECT ON SOD ACTIVITY

SODACTIVITI			
Factor	SOD (u.mg <sup>-1</sup> protein)		
Irrigation (I)			
Normal	10/2.21		
irrigation(I1)	1263.2 b		
Drought stress(I2)	2289.9 a		
Cultivar (C)			
$Zarfam(C_1)$	2010 a		
Okapi (C <sub>2</sub> )	1865 b		
Opera (C <sub>3</sub> )	1454 с		
Selenium(g/ha)			
0 (S0)	1528 с		
16 (S1)	1829 b		
21 (S2)	1973 a		

Similar letters in each column shows non-significant difference according to Duncan's Multiple Range Test

Interaction effect of Irrigation and cultivar showed drought stress condition in Zarfam with 2784 u.mg<sup>-1</sup>protein gained to maximum SOD activity and normal irrigation in Opera located at the lowest part of the Average comparison table (Table III). Drought stress may also lead to stomata closure, which reduces CO2 availability in the leaves and inhibits carbon fixation. This exposes the chloroplast to excessive Excitation

energy, which in turn could increase the generation of free radicals and induce oxidative stress [6].

TABLE III
MEAN COMPARISON INTERACTION EFFECT OF IRRIGATION AND CULTIVARS
EFFECT ON SOD

Factor		SOD (u.mg <sup>-1</sup> protein)
Irrigation	Cultivar	
Normal irrigation	Zarfam	1237 cdef
	Okapi	1438 bc
	Opera	1115 f
Drought stress	Zarfam	2784 cde
	Okapi	2293 def
	Opera	1793 f

Similar letters in each column shows non-significant difference according to Duncan's Multiple Range Test

In the case of Irrigation and selenium, Interaction effect of drought stress condition and 21 g/ha selenium application with 2620 u.mg<sup>-1</sup>protein gained to highest SOD activity (Table IV). The enzymatic antioxidant system is one of the protective mechanisms including superoxide dismutase (SOD: EC 1.15.1.1), which can be found in various cell compartments and it catalyses the isproportion of two  $O2\bar{L}$  radicals to  $H_2O_2$  and  $O2\bar{L}$  [5]–[14].

TABLE IV
MEAN COMPARISON INTERACTION EFFECT OF IRRIGATION AND SELENIUM
EFFECT ON SOD

Factor		SOD (u.mg <sup>-1</sup> protein)
Irrigation	Selenium (g/ ha)	
Normal irrigation	0	1157 e
	16	1307 d
Drought stress	21	1325 d
	0	1899 с
	16	2351 b
	21	2620 a

Similar letters in each column shows non-significant difference according to Duncan's Multiple Range Test

Interaction effect of selenium and cultivar showed Zarfam has high potential for SOD production and increase in selenium spraying to 21 g/ha helps this matter (Table V). Selenium is chemically similar to sulphur, this may cause a non-specific replacement of S by selenium in proteins and other sulphur compounds [11]. There indicates that it can also play a positive biological role in higher plants [3]. Selenium could increase the tolerance of plants to

UV-radiation induced oxidative stress, delay senescence, and promote the growth of ageing seedlings [22]. After study on 3 experimental factor observed that drought stress 21 g/ha selenium spraying in Zarfam variety with 3146 u.mg<sup>-1</sup> protein gained to maximum SOD activity. Plant growth promoted by selenium is due to the increased starch accumulation in chloroplasts [13].

TABLE V
MEAN COMPARISON INTERACTION EFFECT OF VARIETY AND SELENIUM ON
SOD

Facor	300	SOD (u.mg <sup>-1</sup> protein)
Cultivar	Selenium (g/ ha)	
Źarfam	0 16 21	1749 d 2057 b 2224 a
Okapi	0 16 21	1587 ef 1940 c 2069 b
Opera	0 16 21	1248 g 1490 f 1625 e

Similar letters in each column shows non-significant difference according to Duncan's Multiple Range Test

Interaction effect of irrigation, cultivar and selenium on SOD activity had significant difference at %5 level of probability (Table I). The results showed that drought stress and 21 g/ha foliar application of Se in Zarfam variety with 3146 u.mg<sup>-1</sup>protein gained to highest SOD activity and normal irrigation and non Selenium spraying in Opera cultivar with 1042 u.mg<sup>-1</sup>protein produced lowest SOD activity (Table VI). It seems that in the case of SOD activity in varieties there were positive correlation to selenium uses and 21 g/ha of selenium application was more effective than 16 g/ha. Recently it has been shown that selenium can regulate the water status of plants under conditions of water deficiency and thereby performs its protective effect [7]. Selenium can alleviate oxidative stress in chloroplasts. The responses of plant to selenium supplementation were investigated by monitoring chlorophyll fluorescence and the transcription of antioxidative enzymes [15]. When selenium was applied to Trigonella foenum-graecum seedlings, mitochondrial oxygen uptake increased concomitantly with enhanced mitochondrial SOD (superoxide dismutase) activity [19].

The result shows that selenium as foliar application can improve yield under conditions of drought stress and it can be suggested for these lands in arid and semi arid regions [23].

TABLE VI MEAN COMPARISON INTERACTION EFFECT OF IRRIGATION, CULTIVARS AND SELENIUM ON SOD

	Factor		SOD (u.mg <sup>-1</sup> protein)
Irrigation	Variety	Selenium	
Normal	Zarfam	g/ ha 0	1124 cdef
irrigation		16	1283 bc
		21	1303 f
	Okapi	0	1305 ab
		16	1487 a
		21	1521 ef
	Opera	0	1042 bcd
		16	1153 ab
		21	1152 f
Drought	Zarfam	0	2375 a
stress		16	2831 cde
		21	3146 def
	Okapi	0	1868 f
		16	2393 ab
		21	2617 a
	Opera	0	1458 ef
		16	1828 bcd
		21	2098 cdef

Similar letters in each column shows non-significant difference according to Duncan's Multiple Range Test

### REFERENCES

- [1] R.L. Foot, Comparative Responses of Field Grown Crops to Phosphate Concentrations in Soil Solutions. In: Stress Physiology in Crop Plants, Mussell, H. and R.C. Stapley (Eds.).Wiley,NewYork,2001,pp.81-106.URL: www.publish.csiro. au/?act=view file&file id=AR9760479.
- [2] C.H Foyer, M. Lelandais and K.J. Kunert, Photooxidative stress in plants. Physiol. Plant,1992,vol 92 ,pp.696-717. http://cat.inist.fr/?aModele=afficheN&cpsidt=3339390.
- [3] M.Germ, I. Kreft I, and J. Osvald, Influence of UV-B exclusion and selenium treatment on photochemical efficiency of photosystem II, yield and respiratory potential in pumpkins (Cucurbita pepo L.). Plant Physiol. Biochem, 2005, vol. 43, pp. 445-448.
- [4] H.Hartikainen , T. Xue, and V.Piironen, Selenium as an antioxidant and pro-oxidant in ryegrass. Plant Soil, 2000, vol .225,pp. 193-200.
- [5] A. Hegedus, S. Erdei, and G. Horvath, Comparative studies of H<sub>2</sub>O<sub>2</sub> detoxifying enzymes in green and greening barley seedlings under cadmium stress. Plant Sci,2001, vol. 160: 1085-1093. DOI: 10.1016/S0168-9452(01)00330-2.
- [6] S.M. Johnson, S.J. Doherty, and R.R.D. Croy, Biphasic superoxide generation in potato Tubers: A self amplifying response to stress. Plant Physiol,2003, vol 13,pp. 1440-1449. http://www.plantphysiol.org/cgi/content/abstract/1 31/3/1440.
- [7] V.V. Kuznetsov , V.P. Kholodova , V.V. Kuznetsov B.A. Yagodin , Selenium regulates the water status of plants exposed to drought. Dokl. Biol. Sci,2003,vol.390,pp.266-268.
- [8] J.Mwanamwenge, S.P. Loss, K.H.M. Siddique, and P.S. Cocks, Effect of water stress during floral initiation, flowering and podding on the growth and yield of faba bean (Vicia faba L.). Eur. J. Agron, 1999,vol. 11,pp.1-11. DOI: 10.1016/S1161-0301(99)00003-9.

- [9] M.Nasri, M.Khalatbari, H.Zahedi, F.Paknejad, and H.R. Tohidi Moghadam, Evaluation of Micro and Macro Elements in Drought Stress Condition in Cultivars of Rapeseed (Brassica napus L.). American Journal of Agricultural and Biological Sciences, 2003,vol.3 (3),pp.579-583
- [10] G.Noctor, and C.H. Foyer, Ascorbate and glutathione: Keeping active oxygen under control. Annu. Rev. Plant Physiol. Plant Mol. Biol,1998, vol.49,pp 249-279. PMID: 15012235.
- [11] J,Nowak, K.Kaklewski, M.Ligocki , Influence of selenium on oxidoreductive enzymes activity in soil and in plants. Soil Biol. Biochem, 2004, vol. 36, pp. 1553-1558.
- [12] S.Ouchi, A.Nishikawa, and E.Kameda, Soilimproving effect of a super-aterabsorbentpolymer II. Evaporation, leaching of salts and growth of vegetables. Jap. J. Soil Sci. Plant Nutr,1990,Vol.61,pp.606-613. http://www.fao.org/agris/search/display.do?f=/1994/v2007/JP9306919.xml;JP9306919
- [13] A.Pennanen, T.Xue, H.Hartikainen, Protective role of selenium in plant subjected to severe UV irridiation stress. J. Appl. Bot,2002,vol.76,pp. 66-76
- [14] J.G. Scandalios, Oxygen stress and superoxide dismutase. Plant Physiol,1993,vol.101,pp.712-726.
- http://www.pubmedcentral.nih.gov/articlerender.fc gi?artid=158641.
  [15] M.Seppanen , M. Turakainen, H.Hartikainen, Selenium effects on oxidative stress in potato . Plant Science,2003,vol. 165 ,pp.311-/319.
- [16] A.Shalata, V. Mittova, M. Volokita, M. Guy, and M. Tal, Response of the cultivated tomato and its wild salt-tolerant relative Lycopersicon pennellii to salt-dependent oxidative stress: The root antioxidative system. Physiol. Plant,2001,vol. 112: 487-494. DOI: 10.1034/j.1399-3054 2001 1120405 x
- [17] N.Smirnoff, The role of active oxygen in the response to water deficit and desiccation. New Phytol,1993,vol.125,pp. 27-58. DOI: 10.1111/j.1469-8137.1993.tb03863.x.
- [18] H.Soleimanzadeh, D.Habibi, M.R. Ardakani, F.Paknejad, and F.Rejali, Effect of Potassium Levels on Antioxidant Enzymes and Malondialdehyde Content under Drought Stress in Sunflower (Helianthus annuus L.). American Journal of Agricultural and Biological Sciences, 2010, vol. 5 (1), pp. 56-61.
- [19] M. Sreekala, T.R. Santosh, and K. Lilitha, Oxidative stress during selenium deficiency in seedlings of Trigonella foenum-graecum and mitigation by mimosine. Part I. Hydroperoxide metabolism, Biol. Trace Element Res, 1999, vol. 70, pp. 193-207.
- [20] H.Tapiero ,D.M, Townsend, K.D.Tew, Dossier: Oxidative stress pathologies and antioxidants: The antioxidant role of selenium and seleno-compounds. Biomed. Pharmacoth, 2003, vol. 57, pp. 134-144.
- [21] H.R.Tohidi-Moghadam, A.H. Shirani- Rad, G.Nourmmohamadi, D.Habibi, M.Mashhadi-Akbar-Boojar, Effect of Super Absorbent Application on Antioxidant Enzyme Activities in Canola (Brassica napus L.) Cultivars under Water Stress conditions. American Journal of Agricultural and Biological Sciences ,2009,vol.4 (3): 215-223.
- [22] T.L. Xue, H.Hartikainen, V.Piironen, Antioxidative and growth-promoting effects of selenium on senescing lettuce. Plant Soil, 2001, vol. 237, pp. 55-61.
- [23] H.Zahedi, G. Noormohammadi, A. H Shirani-Rad, D. Habibi , M. Mashhadi Akbar Boojar, Effect of Zeolite and Foliar Application of Selenium on Growth, Yield and Yield Component of Three Canola Cultivar under Conditions of Late Season Drought Stress . Not Sci Biol, 2009,vol.1 (1) pp.73-80.