# Influence of Ammonium Concentration on the Performance of an Inorganic Biofilter Treating Methane

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**Abstract**—Among the technologies available to reduce methane emitted from the pig industry, biofiltration seems to be an effective and inexpensive solution. In methane (CH<sub>4</sub>) biofiltration, nitrogen is an important macronutrient for the microorganisms growth. The objective of this research project was to study the effect of ammonium (NH<sub>4</sub><sup>+</sup>) on the performance, the biomass production and the nitrogen conversion of a biofilter treating methane. For NH<sub>4</sub><sup>+</sup> concentrations ranging from 0.05 to 0.5 gN-NH<sub>4</sub><sup>+</sup>/L, the CH<sub>4</sub> removal efficiency and the dioxide carbon production rate decreased linearly from 68 to 11.8 % and from 7.1 to 0.5 g/(m<sup>3</sup>-h), respectively. The dry biomass content varied from 4.1 to 5.8 kg/(m<sup>3</sup> filter bed). For the same range of concentrations, the ammonium conversion decreased while the specific nitrate production rate increased. The specific nitrate production rate presented negative values indicating denitrification in the biofilter.

*Keywords*—Methane, biofiltration, pig, ammonium, nitrification, denitrification.

# I. INTRODUCTION

WITH a total production of 109 million tons of pig meat in 2010 [1], the world pork industry was also responsible for water, air and soil pollution [2]. Among the compounds responsible for air pollution, this agricultural sector released volatile fatty acids, ammonia (NH<sub>3</sub>), hydrogen sulphide (H<sub>2</sub>S) and greenhouse gases (GHG) such as carbon dioxide (CO<sub>2</sub>) and methane (CH<sub>4</sub>) [3]. In Canada (2009), CH<sub>4</sub> emissions represented 13% of the total GHG emissions (690 Mt eq.  $CO_2$ ) which corresponds to 90 Mton eq.  $CO_2[4]$ . In 2004, the world CH<sub>4</sub> anthropogenic emissions represented around 6.9 Gton eq. CO<sub>2</sub>[5]. With a heat of combustion of 890 kJ/mol (25 °C, 1 atm) [6], CH<sub>4</sub> is an interesting compound produced by anaerobic digestion of organic matter [3]. However, CH<sub>4</sub> emissions, even if they are lower than CO<sub>2</sub> emissions, are not negletable in terms of global warning because CH<sub>4</sub> has a global warming potential 25 times higher than CO<sub>2</sub> over a period of 100 years [7].

Even if CH<sub>4</sub> can theoretically be thermally oxidized, the latter requires a minimal CH<sub>4</sub> concentration in air ranging from 5 to 15% (v/v) [8]. In case of  $CH_4$  emitted from slurry storage, the concentrations are generally lower than 3% (v/v), which is not enough to use thermal oxidation [9]. On the other hand, several studies have shown that low CH4 concentrations can be treated effectively and relatively non-expensively by biofiltration [9]. In order to increase the biofilter performance, some parameters must be controlled such as moisture, temperature and nutrients [10]. Among the nutrients, microorganisms require nitrogen because it represents up to 14% of dry cell weight [11]. Usually, nitrogen is supplied to inorganic bed biofilters as a form of nitrate (NO3) [12] because ammonium  $(NH_4^+)$  had a negative effect (inhibiting potential on CH<sub>4</sub> oxidation) on methanotrophic bacteria in soil studies [13, 14], but also had a positive effect (stimulation of CH<sub>4</sub> oxidation) in other soils studies [15].

The objective of this study was to test the effect of  $NH_4^+$  concentration in the nutrient solution of the performance of an inorganic packed bed biofilter treating CH<sub>4</sub>. The performance of the biofilter was determined by analyzing the carbon and the nitrogen balance.

#### II. MATERIALS AND METHODS

Fig. 1 presents the inorganic packed bed biofilter used for the experiments. The biofilter was a Plexiglas cylinder with an inlet diameter of 15 cm, divided into 3 sections. The biofilter was packed with an inorganic material for a total bed height of 1 m (volume of 18 L). The exact nature of the filter bed cannot be revealed for confidential reasons.

A mixture of pure CH4 (Praxair) and compressed air containing oxygen  $(O_2)$  was fed at the bottom of the biofilter and the treated air was released at the top. In order to avoid filter bed desiccation, the air mixture was previously saturated with water by passing through a humidification column. A nitrate salts medium (NMS) was used to supply nutrients and moisture to the filter bed [17]. At the top of the biofilter, the nutrient solution was fed (1.5 L; once a day) while the leachate was collected at the bottom of the biofilter. Concurrently, NO3<sup>-</sup> (as sodium nitrate) concentration was decreased by 0.05 gN-NO<sub>3</sub>/L increasing steps and NH<sub>4</sub><sup>+</sup> (as ammonium carbonate)concentration was increased in order to keep the total nitrogen concentration in the nutrient solution at 0.5 gN/L. The NO<sub>3</sub> concentration was decreased from 0.45 to 0 gN-NO<sub>3</sub><sup>-</sup>/L while the  $NH_4^+$  concentration was increased from 0.05 to 0.5 gN-NH<sub>4</sub><sup>+</sup>/L.

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Fig. 1 Biofilter set up for the biofiltration of CH<sub>4</sub>

Suspended biomass contained in leachate samples was removed using filter paper. Ionic chromatography (Dionex ICS-1000, Canada) was employed to determinate the concentrations of  $NH_4^+$ ,  $NO_3^-$  and  $NO_2^-$  in leachate and nutrient solution [18]. The weight difference of the packing material sample dried at 105 °C and calcined at 500 °C was used to determinate the dry biomass concentration in the packed bed [19].

At the bottom of the biofilter, the CH<sub>4</sub> concentration and air flow rate were respectively set at 1500 ppmv (0.15% v/v) (inlet load of 10 g/(m<sup>3</sup>-h)) and 3 L/min. At each sample port, a total hydrocarbon analyser equipped with a continuous flame ionisation detector (Horiba model FIA-510, USA) was utilized to measure the CH<sub>4</sub> concentration in the gas phase. A gas analyser detector (Ultramat 22P, Simens, Germany) was also employed to measure the CO<sub>2</sub> concentration in the gas phase. Table I summarizes the main parameters considered to evaluate the performance of the biofilter. The theoretical dry biomass production rate (DBR), listed in Table I, was used to evaluate the theoretical dry biomass production. This parameter is evaluated by means of a molar balance of  $CH_4$  and  $CO_2$ .

### III. RESULTS AND DISCUSSION

#### A. Biofilter Performance

Fig. 2 presents the methane removal efficiency (CH<sub>4</sub>-RE) and the  $P_{CO_2}$  as a function of the NH<sub>4</sub><sup>+</sup> concentration in the nutrient solution. For NH<sub>4</sub><sup>+</sup> concentrations from 0.05 to 0.5 gN-NH<sub>4</sub><sup>+</sup>/L, the CH<sub>4</sub>-RE decreased linearly from 68 to 12%. For NH<sub>4</sub><sup>+</sup> concentrations ranging from 0.2-0.25 gN-NH<sub>4</sub><sup>+</sup>/L, the CH<sub>4</sub>-RE decreased quickly from 50 to 24%. For NH<sub>4</sub><sup>+</sup> concentrations ranging from 0.05 to 0.15 gN-NH<sub>4</sub><sup>+</sup>/L, the P<sub>CO<sub>2</sub></sub> increased from 7.1 to 12.4 g/(m<sup>3</sup>-h) and from 0.15 to 0.5

# gN-NH<sub>4</sub><sup>+</sup>/L, the $P_{CO_2}$ decreased from 12.4 to 0.5g/(m<sup>3</sup>-h).

The fact that CH<sub>4</sub>-RE decreased with the NH<sub>4</sub><sup>+</sup> concentration shows the effect of NH<sub>4</sub><sup>+</sup> on the populations of methanotrophic bacteria present in the biofilter. Many studies have shown that NH<sub>4</sub><sup>+</sup> reduces the CH<sub>4</sub> oxidation rate in soil [20, 21, 22, 23], compost [24] and biofilters [18, 25, 26]. For example, in paddy soil, for CH<sub>4</sub> inlet concentrations of 1500 ppmv, Cai and Mosier [20] found that for an increase of NH<sub>4</sub><sup>+</sup> concentration from 0 to 0.05 mgN-NH<sub>4</sub>/kg soil, the CH<sub>4</sub> oxidation rate decreased from 338 to 166 ngC-CH<sub>4</sub>/(g soil-h). In the present study, the decrease of CH<sub>4</sub>-RE from 68 to 12% (-83%) was more important because, in soil, the nitrifying bacteria are already present, which reduced the NH4<sup>+</sup> concentration in the filter bed. This could also mean that a  $NH_4^+$  concentration of 0.5 gN- $NH_4^+/L$  has more negative effect on CH<sub>4</sub> oxidation than the NH<sub>4</sub><sup>+</sup> concentration used by Cai and Mosier [20](0.05 mgN-NH<sub>4</sub>/kg soil).

Between 0.05 and 0.15 gN-NH<sub>4</sub><sup>+</sup>/L, the  $P_{CO_2}$  increased from 7.1 to 12.4 g/(m<sup>3</sup>-h) (+76%) even if CH<sub>4</sub>-RE decreased from 68 to 57% (-16%). This may means that less carbon was used to produce biomass inducing a lower methanotrophic

	MAIN PARAMETERS USED TO EVALUATE THE BIOFILTER PERFORMANCE	
Parameters	Equations	Units
Methane removal efficiency (CH <sub>4</sub> -RE)	$CH_4 - RE = \frac{C_{gin} - C_{gout}}{C_{gi}}$	Dimensionless
Carbon dioxide production rate ( $P_{CO_2}$ )	$P_{CO_2} = \frac{Q^*(C_{dout} - C_{din})}{V}$	g/(m <sup>3</sup> -h)
Theoretical dry biomass production rate (DBR)	$DBR = \left[\frac{\left(C_{gin} - C_{gout}\right)}{W_{CH_4}} - \frac{\left(C_{dout} - C_{din}\right)}{W_{CO_2}}\right] \left(\frac{Q \cdot W_B}{V}\right)$	g biomass/(m <sup>3</sup> -h)
Nitrate production rate ( $P_{NO_3}$ )	$P_{NO_{3}} = \frac{(NO_{3in}^{-} - NO_{3out}^{-}) \cdot Q_{NS}}{V * DB}$	gN/(g biomass-h)

TABLE I Main Parameters used to Evaluate the Biofilter Performance

a:  $C_{gin}$  and  $C_{gout}$  are the inlet and outlet concentrations of  $CH_4$  (g/m<sup>3</sup>); Q is the air flow; V is the volume of the biofilter (0.018 m<sup>3</sup>);  $C_{din}$  and  $C_{dout}$  are the inlet and outlet concentrations of carbon dioxide (g/m<sup>3</sup>);  $W_{CH_4}$ ,  $W_{CO_2}$  and  $W_B$  are the molecular weights of  $CH_4$ ,  $CO_2$  and biomass produced (g/mol),

assuming an empirical formula of  $C_5H_7NO_2$  for biomass with an average value of 113 g/mol [16];  $NO_{3in}^-$  and  $NO_{3out}^-$  are the concentration of  $NO_3^-$  in the nutrient solution and the leachate, respectively (gN/L);  $Q_{NS}$  is the flow of nutrient solution (L/h); DB is the dry biomass in the filter bed (g biomass/(m<sup>3</sup> filter bed)).

activity, explaining the CH<sub>4</sub>-RE decrease. For NH<sub>4</sub><sup>+</sup> concentrations higher than 0.15 gN-NH<sub>4</sub><sup>+</sup>/L, the  $P_{CO_2}$  followed the same tendency than the CH<sub>4</sub>-RE. Between 0.20 and 0.25 gN-NH<sub>4</sub><sup>+</sup>/L, the quickly decrease of  $P_{CO_2}$  from 9.0 to 4.7 (-48%) confirms the assumption that for this concentration range, a major change in bacteria population occurred in the biofilter as observed in a previous study [19].



Fig. 2  $CO_2$  production rate (**•**) and  $CH_4$  removal efficiency ( $\circ$ ) as a function of the  $NH_4^+$  inlet concentration in the nutrient solution

#### B. Biomass

Fig. 3 presents the average dry biomass content (DB) and the theoretical dry biomass production rate (DBR) as a function of the inlet  $NH_4^+$  concentration in the nutrient solution. For  $NH_4^+$  concentrations ranging from 0.05 to 0.5 gN- $NH_4^+/L$ , the DB decreased with the  $NH_4^+$  concentration and varied from 5.8 to 2.5kg/m<sup>3</sup> filter bed. The DBR decreased also with the  $NH_4^+$  concentration and followed a logarithmic tendency with values ranging from 30 to 5 g/(m<sup>3</sup>h).

The decrease of DBR with the  $NH_4^+$  concentration was also observed by Wilshusen et al. [24]. In order to explain this phenomena, the authors hypothesized that the exopolymeric substances could serve "as a carbon cycling mechanism for type I" methanotrophic bacteria. The fact that DBR decreased could also indicate that more carbon was transformed into  $CO_2$ , which explains the decrease of CH-RE observed in Fig. 2 as less new biomass was formed.

The fact that the DB (linear) followed a different tendency than DBR (logarithmic) indicates that some microorganisms other than methanotrophic bacteria (like denitrifying and nitrifying bacteria) can generate biomass. However, a visual inspection of the biofilter shows a decrease of the biomass in the filter bed which may means that the biomass produced by other microorganisms may be more soluble in water. The dry biomass content is also influenced by the amount of biomass washed out of the filter bed at each daily watering. The decrease of DB observed in the filter bed (-57%) was lower than the DBR(-83%). As a consequence, for the NH<sub>4</sub><sup>+</sup> concentrations tested, less biomass would be lost in the leacheate as the NH<sub>4</sub><sup>+</sup> concentration increased.



Fig. 3 Dry biomass content ( $\diamond$ ) and theoretical dry biomass production rate (**•**) as a function of the NH<sub>4</sub><sup>+</sup> inlet concentration in the nutrient solution

#### C.Nitrogen Conversion

Fig. 4 presents the  $NH_4^+$  conversion and the specific  $NO_3^-$  production rate  $(P_{NO_3})$  as a function of the  $NH_4^+$  inlet concentration in the nutrient solution. For  $NH_4^+$  concentrations in the nutrient solution ranging from 0.05 to 0.5 gN- $NH_4^+/L$ , the  $NH_4^+$  conversion decreased linearly from 48 to 26 % while the  $P_{NO_3}$  increased from -0.01 to 0.16 gN/(m<sup>3</sup>-h).



Fig. 4 Ammonium conversion (♦) and specific nitrate production rate (Δ) as a function of the NH<sub>4</sub><sup>+</sup> inlet concentration in the nutrient solution

The fact that the NH<sub>4</sub><sup>+</sup> conversion decreased with the NH<sub>4</sub><sup>+</sup> concentration could be due to the decrease of CH<sub>4</sub>-RE (Fig. 2). In fact, the increase of CH<sub>4</sub> concentration could lead to the decrease of NH<sub>4</sub><sup>+</sup> conversion as CH<sub>4</sub> is an inhibitor of nitrifying bacteria [27]. Moreover, the increase of CH<sub>4</sub> concentration in the biofilter could also lead to changes of number and kind of microorganisms specific to NH<sub>4</sub><sup>+</sup> conversion deacrease. The P<sub>NO3</sub> presented some negative values at 0.05 and 0.10 gN-NH<sub>4</sub><sup>+</sup>/L of -0.01 and -0.02gN-NO<sub>3</sub><sup>-</sup>/(g biomass-

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h), respectively. This indicated that there was a consumption of  $NO_3^-$  by methanotrophic bacteria or a denitrification.

# IV. CONCLUSION

Increasing the NH<sub>4</sub><sup>+</sup> concentration in the nutrient solution reduced the performance of an inorganic biofilter treating CH<sub>4</sub> at an inlet concentration of 1500 ppmv, as follows: the CH<sub>4</sub>-RE, the  $P_{NO_3}$  and the dry biomass content decreased respectively from 68 to 12 %, from 7.1 to 0.5 g/(m<sup>3</sup>-h) and from 5.8 to 4.1 kg/m<sup>3</sup> filter bed. For the same range of concentrations, the NH<sub>4</sub><sup>+</sup> conversion also decreased from 48 to 26% whereas the  $P_{NO_3}$  increased from -0.01 to 0.16 gN/(m<sup>3</sup>-h) which suggests that denitrification occurred. This study shows that the nature and the concentration of the macronutrients (nitrogen) present in the nutrient solution are important for the performance of an inorganic biofilter treating CH<sub>4</sub>.

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