# The Effect of Acute Toxicity and Thyroid Hormone Treatments on Hormonal Changes during Embryogenesis of *Acipenser persicus*

Samaneh Nazeri, Bagher Mojazi Amiri, Hamid Farahmand

Abstract-Production of high quality fish eggs with reasonable hatching rate makes a success in aquaculture industries. It is influenced by the environmental stimulators and inhibitors. Diazinon is a widely-used pesticide in Golestan province (Southern Caspian Sea, North of Iran) which is washed to the aquatic environment (3 mg/L in the river). It is little known about the effect of this pesticide on the embryogenesis of sturgeon fish, the valuable species of the Caspian Sea. Hormonal content of the egg is an important factor to guaranty the successful passes of embryonic stages. In this study, the fate of Persian sturgeon embryo to 24, 48, 72, and 96-hours exposure of diazinon (LC50 dose) was tested. Also, the effect of thyroid hormones (T3 and T4) on these embryos was tested concurrently or separately with diazinon LC 50 dose. Fertilized eggs are exposed to T3 (low dose: 1 ng/ml, high dose: 10 ng/ml), T4 (low dose: 1 ng/ml, high dose: 10 ng/ml). Six eggs were randomly selected from each treatment (with three replicates) in five developmental stages (two cell- division, neural, heart present, heart beaten, and hatched larvae). The possibility of changing T3, T4, and cortisol contents of the embryos were determined in all treated groups and in every mentioned embryonic stage. The hatching rate in treated groups was assayed at the end of the embryogenesis to clarify the effect of thyroid hormones and diazinon. The results indicated significant differences in thyroid hormone contents, but no significant differences were recognized in cortisol levels at various early life stages of embryos. There was also significant difference in thyroid hormones in (T3, T4) + diazinon treated embryos (P<0.05), while no significant difference between control and treatments in cortisol levels was observed. The highest hatching rate was recorded in HT3 treatment, while the lowest hatching rate was recorded for diazinon LC50 treatment. The result confirmed that Persian sturgeon embryo is less sensitive to diazinon compared to teleost embryos, and thyroid hormones may increase hatching rate even in the presence of diazinon.

Keywords-Persian sturgeon, diazinon, thyroid hormones, cortisol, embryo.

# I. INTRODUCTION

**D**IAZINON (O,O-Diethyl O-[4-methyl-6-(propan-2yl)pyrimidin-2-yl] phosphorothioate) is a widely organophosphate pesticide used in rice paddy fields of Golestan province, Iran to control insects [30]; then, it is washed to the aquatic environment [20], [28], [34], i.e. enters in the rivers leading to the Caspian Sea [41], [46]. There are evidences on the presence of diazinon in the rivers of Golestan province such as Qara-su River (18.6-22.4 mg/l) and Gorgan-rud River (6.74-6.89 mg/l) [49].

There are some environmental factors that affect the rate of diazinon degradation. Low temperature, low moisture, high alkalinity, and lack of adequate microbial degraders for more than six months cause decrease in diazinon degradation [15], [20], [28]. It also enters into the running water based on the time, concentration, frequency of use, and also rainfall [56]. Diazinon degradation through hydrolysis, photolysis, volatilization, and microbial metabolism occurs in small amount to break down the chemical [2], so it is persistent and makes high contamination. Vast distribution of diazinon in aqua ecosystems affects non-target aquatic organisms such as fish.

Sub-lethal dose of diazinon varies from one species to another [5], [23] and resulted in inhibition of acetylcholinesterase activity [43]. Organophosphates also hamper neurotransmitters activity which then affect thyroid hormone function [3], [36]; it also alters thyroid hormones and cortisol concentrations in fish [31]. It is shown that fishes are the most susceptible to the toxic during the developmental process of embryos and larvae [22], [28], [33].

Maternal hormones such as thyroid hormones (THs) and cortisol are important in the regulation of growth, development, sex determination, and survival of fish embryos and larvae [11], [12], [37], [38]. Usually during the development, the levels of maternal hormones decrease, and endocrine organs start to differentiate. Probably, artificial increase in the levels of maternal hormones will be a key factor in larviculture [53]. Studies showed that THs in the fish oocyte increased by T3 injection to the female breeders [18], and positive effects of increase in maternal T3 level have been proved in larval survival [7], [12], [13].

The present study was initiated to determine acute toxicity of diazinon in the embryos of Persian sturgeon, *Acipenser persicus*; one of the commercial and valuable sturgeon fishes in the Caspian Sea [40], [44]. Persian sturgeon is a suitable species for the study because of long term exposure to toxins in their ecosystems.

There are internal and external factors to affect the quality of the fish eggs [10]. This study is designed *in vitro* to evaluate the effect of sublethal diazinon and also THs on some embryonic hormonal contents in order to contribute the knowledge of the early life stages of fish. The critical period that ultimately makes the fate of an embryo.

Samaneh Nazeri is with the Department of Fisheries and Environmental sciences, Faculty of Natural Resources, University of Tehran, 31587-77871 Karaj, Iran (e-mail: s\_nazeri@ut.ac.ir).

# II. MATERIALS AND METHODS

# A. Collection of the Eggs and Artificial Fertilization

Persian sturgeon fish embryos used for acute toxicity tests were obtained from autumn brood stock program of Shahid Marjani Hatchery Complex near Gorgan city (North-Eastern of Iran). All samples were received from artificially fertilized oocytes of one wild female (24 kg in weight and 152 cm length) and sperm of three wild males.

# **B.** Experimental Treatment

Diazinon (95% purity) was purchased from Qingdao Hisigma chemicals Co. Ltd (China). The nominal treatment concentrations (0, 2, 4, 6, 8 mg/L) were used for any of different embryonic stages [6]. The medium was the sterile of well water used at Shahid Marjani Hatchery Complex.

After the second egg cleavage, the fertilized eggs were separated from unfertilized, parthenogenesis, and poly sperm eggs. 300 fertilized eggs (60 for each treatment) were used for the experiment. Samples were transferred to the central lab, University of Tehran within six hours.

# C. Acute Toxicity Test

Embryos were incubated in six well-culture dishes (SPL, Korea) with no or different concentrations of diazion and put into CO2 incubator under 16 °C. Incubation period lasted for 24, 48, 72, and 96 hours. It was semi-static method, and the culture media were changed daily with consideration of the mortalities. The procedure was arranged according to OECD guideline for fish embryo toxicity (FET) test [26].

# D. Hormonal Assays

The second part of the test was carried out after determining the LC<sub>50</sub> value. The fertilized eggs were obtained as described before and transported to the laboratory within six hours. There were 10 groups with three replicates each group in six well-culture dishes. One group was just in medium as control, one group was toxin treatment in medium as LC50 dose, two groups were low dose (LD) of T3 (tri-iodothyronine) and T4 (thyroxine) (1 ng/ml) in medium, two groups were high dose (HD) of T3 and T4 (10 ng/ml) in medium, two groups with the mixture of LC50 diazinon and LD of T3 or T4 in medium and two other groups were the mixture of LC50 diazinon and HD of T3 or T4 in medium.

The concentrations for low dose of T3 and T4 treatments were chosen similar to the results of a previous study by [9] for THs in the fertilized eggs of sturgeon fish, and the high dose were selected more than the result of [39] for THs in plasma of juvenile sturgeon fish. There were 90 embryos in each group. The embryos were kept in CO<sub>2</sub> incubator under desired condition until they hatched and removed only for monitoring the developmental stages at defined time by stereomicroscope (10X). The samples for hormonal assays were removed in five embryonic stages (2 cell-division, neurula, heart present, heart beaten and immediately after hatch) to evaluate the hormonal contents in each stage.

# E. THs Analysis

TH extraction was performed according to [52] and [47]. Each three frozen eggs or larvae were homogenized in two volumes of phosphate- buffered saline (PBS; pH=7.5), and the mixtures were sonicated. Then, 0.167 volume of a 10X stock trypsin solution (Inoclon) was added to each tube and was vortexed to release thyroglobulin from thyroid tissues, then incubated in shaking water bath at 38 °C for 1 hour. After that, 2 ml ice-cold ethanol (99%) added to the samples, was vortexed and centrifuged at 3000 rpm for 10 minutes at 4 °C. The supernatant was decanted, and 2 mL ice-cold ethanol (99%) was again added again to the residue, vortexed and centrifuged. The supernatant was extracted and pooled to the previous one. Finally, it was evaporated overnight, then was frozen at -20 °C until later hormone assay.

# F. Cortisol Analysis

Cortisol extraction was followed by [35]. Every three embryos were homogenized in 800 µl distilled water. The tubes were vortexed. 8 ml diethyl ether was added to the samples and then vortexed vigorously for 30 seconds. Diethyl ether was applied for the second time about 4 ml and they were vortexed again. Samples centrifuged at 2100 g for 5 minutes at 4 °C. Supernatants were evacuated to other tubes. At the end of the procedure, they were reconstituted with phosphate- buffered saline containing gelatin (PBSG).

# G. Enzyme Immunoassay of hormones

T3, T4 (Pishtaz Teb, Iran) and cortisol (DiaMetra, Italy) enzyme immunoassay tests were performed in duplicate using up to 50, 25, and 20 µL, respectively for each sample. The OD values of standards and experimental samples were read within 10 minutes using a microplate reader (BioTek, ELx808) with a 450-nm filter. A standard curve was drawn ranging from 0 to 20 for T3 or T4 and 0 to 40 ng/ml for cortisol. The coefficient in linear range of the curves was almost 0.8 for all of them to calculate the hormone concentrations.

# H. Statistical Analysis

After 96 hours, the mortalities were assessed by statistical package SPSS 16.0 (Chicago, IL). The data were subjected to Probit Analysis Statistical Method to determine the 96h LC<sub>50</sub>.

The data obtained, expressed as means ± SE, were statistically analyzed by multivariate analysis and significant differences were detected. Duncan multiple range test was performed to determine significant differences among groups (*P*<0.05).

## **III. RESULTS**

# A. Determination of Sub-Lethal Dose of Diazinon

The number of dead embryos for diazinon doses of 2, 4, 6, and 8 mg/L were examined for 24, 48, 72, and 96 hours of exposure in Persian sturgeon embryos. The 96LC50 value (95% confidence limits) of water-soluble diazinon of Persian sturgeon fish at early life stage was detected as 3.558 mg/L.

# International Journal of Biological, Life and Agricultural Sciences ISSN: 2415-6612 Vol:11, No:2, 2017

# B. TH Level

All the embryos were used for hormonal extractions. The whole-body T3 concentration of Persian sturgeon ranged from  $8.47\pm1.07$  to  $13.38\pm1.63$  pg/mg in control during embryonic stages (after fertilization until immediately after hatch). T3 content also varied significantly by embryonic development ( $F_{5,120}$ = 18.969, *P*<0.05; Fig. 1). T3 level increased significantly (*P*<0.05) in the low dose of T3 (LT3) and high dose of T3- treated group (HT3) compared to the control and other treated groups. In contrast, LC<sub>50</sub>- treated group showed

decrease in T3 level significantly (F  $_{9,120}$  =18.126, P<0.05; Fig. 1).

The whole-body T4 concentration of Persian sturgeon embryos ranged from 140.40±9.61 to 262.25±45.36 pg/mg in control (after fertilization until immediately after hatch). The highest T4 level was observed in low dose of T4-treated group (LT4) and the lowest one in LC<sub>50</sub>- treated group (F <sub>9,120</sub>= 19.954, *P*<0.05). Like T3, T4 content of the body varied significantly during the embryogenesis (F <sub>5,120</sub>= 198.491, *P*<0.05; Fig. 2).

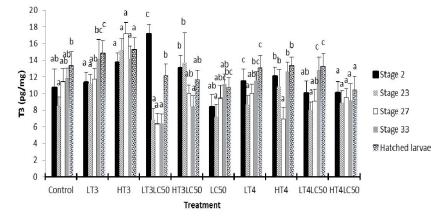


Fig. 1 T3 concentrations (pg/mg) in developing Persian sturgeon embryos following exposure to all treated groups or control. X-axis shows treated groups: LT3: low dose of T3; HT3: high dose of T3; LT3LC50: the mixture of low dose of T3 and LC<sub>50</sub> of diazinon; HDLC50: the mixture of high dose of T3 and LC<sub>50</sub> of diazinon; LC50: only diazinon in the amount of LC<sub>50</sub>; LT4: low dose of T4; HT4; high dose of T4; LT4LC50: the mixture of high dose of T4 and LC<sub>50</sub> of diazinon; HT4LC50: the mixture of high dose of T4 and LC<sub>50</sub> of diazinon; stage23: neurula; stage 27: heart present; stage 33: heart beaten and the larvae immediately after hatching. Data are presented as mean±SEM

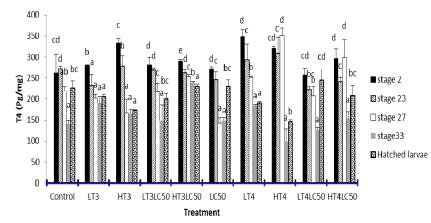


Fig. 2 T4 concentrations (pg/mg) in developing Persian sturgeon embryos following exposure to all treated groups or control

## C. Cortisol Level

Although there was fluctuation during embryogenesis and also an increase in cortisol content after hatching but this increase did not become statistically significant in early development stages of control group. There was also no significant difference found among all treated groups in cortisol level (Fig. 3). The whole-body cortisol content in control was from  $2.4\pm1.22$  to  $3.59\pm1.18$  pg/mg during

embryogenesis (after fertilization until immediately after hatch).

#### D. Hatching Rate

The hatching rate of different treated groups is shown in Fig. 4. The highest hatching rate was recorded in low dose and high dose of T3 and high dose of T4-treated group. The hatching rate revealed significant difference among all treated groups (P=0.000).

# International Journal of Biological, Life and Agricultural Sciences ISSN: 2415-6612 Vol:11, No:2, 2017

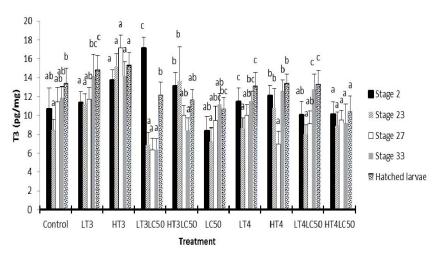


Fig. 3 Cortisol concentrations (ng/mg) in developing Persian sturgeon embryos following exposure to all treated groups or control

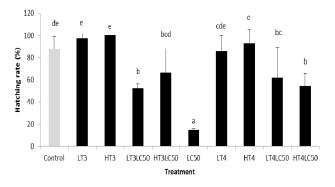


Fig. 4 The hatching rate in Persian sturgeon embryos following exposure to all treated groups or control. Each column represents the mean value of three replicates

# IV. DISCUSSION

# A. Acute Dose of Diazinon

In this study, the LC<sub>50</sub> value of organophosphate pesticide diazinon was calculated for Persian sturgeon embryos for the first time. This value showed that sturgeon embryos are more resistant to the diazinon compared to the other fish embryos as it is observed in common carp, *Cyprinus carpio* with 48h LC<sub>50</sub> value of 0.999 (0.698-1.427) mg/L [6], 96-h LC<sub>50</sub> value for eyed egg and alevins of chinook salmon, *Oncorhynchus tshawytscha* with 545 and 29.5 ppm, respectively [55] and 96-h LC<sub>50</sub> value for European cat-fish fingerling, *Silurus glanis* L. was 14.597 (12.985-16.340) mg/L, 9.710 ppm for young Gambusia [48] and 96-h LC<sub>50</sub> value for Persian sturgeon fish fingerling was 1.8 mg/L [27]. Sturgeon fish embryos have a thick envelope consisting of four distinct layers [16]. It is probably a barrier for low external chemicals diffusion to the inside of eggs.

In this experiment, the sub-lethal dose of diazinon was used to determine the effect of this chemical on sturgeon embryos as the previous studies showed that the presence of Man-built dams especially at Gorganrud and discharges of chemicals to the rivers and estuaries cause the high pollution environment for the fish [4], [32]. The concentration of diazinon is recorded until 3.8 mg/L [49], [50] in the rivers of southern Caspian Sea basin. These higher pesticide concentrations usually happen during spring when the precipitation is low [41], and unfortunately fish restocking performed [1].

The results showed that the all concentrations of media diazinon caused obvious hormonal changes in the whole Persian sturgeon embryo content. THs decreased significantly in diazinon LC<sub>50</sub> treated group, but no significant difference was recorded in cortisol levels. It was reasonable that the lowest hatching rate was pertained to diazinon LC50 treated group. Similar studies confirm decrease of THs against organophosphates [31]. They showed that chlorpyrifos, an organophosphate pesticide, induces decrease in the level of T3 and T4 in Bloch, Heteropneustes fossilis weighing 10±0.5 g embryos. Diazinon decreased T3 and T4 significantly in Caspian roach, Rutilus rutilus fingerlings [30] also in Persian sturgeon fingerlings [27]. The changes of T3 and T4 levels in our study support the interaction of these hormones during early development. Some amounts of diazinon also caused decrease in plasma cortisol level in spotted scat, Scatophagus argus with a mean weight of 137±6 g [24]. Plasma cortisol increased in Persian sturgeon fingerlings during short term exposure of diazinon [27]. Although the developing embryos are more sensitive to endocrine-disrupting chemicals (EDC), but effects on early life stages may not be revealed until adulthood [25]. Sometimes, extremely low-dose exposure of chemicals makes significant effect on developing organisms [57], but it is worth mentioning that effects are often appeared by non-traditional dose-response curves [25], [57], so maybe some doses of lower diazinon exert cortisol alteration.

Higher mortality was recorded in Zebrafish, *Danio rerio* embryos exposed to 3000 µg/L diazinon [42]. Diazinon exposure manifested decrease in hatch success in Medaka, *Oryzias latipes* embryos too [28].

# B. THs-Treated Groups and Control

The possibility of passive diffusion of THs in to eggs may lead to the increase of TH levels in embryos significantly compared to the control. There were no significant changes in cortisol content among T3 and T4 treated groups and the others. The present results demonstrated that the hatching rate increased significantly in low dose and high dose of T3-treated groups and high-dose of T4-treated group. Female brood injection of T3 and maternally transfer to the larvae increases growth and survival [14]. Elevating the T4 level in embryos and larvae probably prevents from heavy mortality in some species [7].

There are many reports of fish eggs THs content such as chum salmon, *Oncorhynchus keta* with 9 ng/g T3 and 15-20 ng/g T4, sockeye salmon, *Oncorhynchus nerka* with 1ng/egg T3 and 6 ng/egg T4, Stripped bass with 4.5 ng/g T3 and 5.1 ng/g Tilapia with 11.4 ng/g T3 and 2.1 ng/g T4, Rabbit fish with 2ng/g T3 and 10-16 ng/g T4, Conger eel, *Conger myriaster* with 0.15 ng/g T3 and 5ng/g T4 [45] and Rassian sturgeon fish, *Acipenser guldenstaedtii* with mean maximum  $0.025\pm0.004$  ng/egg T3 and  $0.34\pm0.09$  ng/egg T4 [9] while in this case study, the value of T3 in control was  $8.47\pm1.07$  to  $13.38\pm1.63$  pg/mg. The value of T4 in control was recorded from  $140.40\pm9.61$  to  $262.25\pm45.36$  pg/mg during early developmental stage. Since these two species of sturgeon fish are so close in phylogenetic, similarity in hormonal content of eggs is expected.

In this study, the cortisol content of control varied from 2.4±1.22 to 3.59±1.18 pg/mg during embryogenesis (after fertilization until immediately after hatch). Data obtained are quite similar to what were recorded by Falahatkar et al., in the same species [21]. Simontacchi et al. expressed that cortisol content of white sturgeon, Acipenser transmontanus differed from 0.73±0.22 to 3.76±0.53 ng/g during early development [51]. Cortisol level showed no significant difference among all treated groups in this study. Falahatkar et al. demonstrated that stress exposure to developing eggs had no effect on wholebody cortisol concentration [21]. Most evidences showed that cortisol and THs activities are synergistic during metamorphism in larvae [19], [29]. It seems that HIP axis is not functional at early stages of sturgeon fish life or probably the synergism of cortisol and THs is species specific bioassay for toxicological assessment. Diazinon decreases ACTH and dbcAMP-stimulated cortisol secretion. Adrenotoxic pattern of diazinon in trout, Oncorhynchus mykiss adreno cells, was presented by Bisson and Hontela [8]. Diazinon probably is less cytotoxic to sturgeon fish during embryogenesis as it is suggested that diazinon at the endocrine-disrupting dose is not cytotoxic to the rat adrenal cells [17].

# C. THs and Acute Toxicity Effects

In this study, T3 content of embryos in the mixed treated groups of  $LT3+LC_{50}$  value of diazinon ( $LT3LC_{50}$  of diazinon) or  $HT3+LC_{50}$  value of diazinon ( $HT3LC_{50}$  of diazinon) was higher compared to  $LT4LC_{50}$  of diazinon and  $HT4LC_{50}$  of diazinon-treated groups, and all of them were higher than  $LC_{50}$  value of diazinon- treated group.

The results showed that T4 content of embryos in the mixed treated groups of  $LT4+LC_{50}$  value of diazinon or  $HT4+LC_{50}$  value of diazinon was higher than  $LC_{50}$  value of diazinon

treated group and also  $LT3LC_{50}$  of diazinon. But, they are lower than control and  $HT3LC_{50}$  of diazinon.

The rate of hatching in hormonal plus  $LC_{50}$  of diazinon treated group was higher than  $LC_{50}$  of diazinon treated group but lower than control and THs-treated groups. Thangavel et al. demonstrated that reduction of T3 in *Sarotherodon mossambicus* under dimecron (an orghanophosphate pesticide) exposure leads to drop in oxygen uptake of fish [54]. It is suggested that decreasing hatching rate in embryo exposed to the toxin might possibly be compensated by adding external THs.

The present results suggest that sturgeon fish breeding in to the diazinon contaminated Caspian Sea basin may alter the embryos and larvae physiologically and the ability of hatch and endanger their survival and ultimately Persian sturgeon fish stocks in the Caspian Sea.

#### ACKNOWLEDGMENT

We wish to express our gratitude to Iranian Fisheries Organization and Shahid Marjani management for supporting this research by donating the eggs.

#### References

- Abdolhay, H. A. and H. B. Tahori (2006). Fingerling production and Release for Stock Enhancement of Sturgeon in the Southern Caspian Sea: an overview. Journal of Applied Ichthyology, 22: 125-131.
- [2] Ahmadi-Mamaqani, Y., N. Khorasani, K. Talebi, S. H. Hashemi, G. Rafiee and F. Bahadori-Khosroshahi (2011). Diazinon fate and toxicity in the Tajan River (Iran) ecosystem. Environmental Engineering Science, 28: 859-868.
- [3] Akhtar, N., S. Kayani, M. Ahmad and M. Shahab (1996). Insecticide-induced Changes in Secretory Activity of the Thyroid Gland in Rats. Journal of Applied Toxicology, 16: 397-400.
- [4] Aladin, N. and I. Plotnikov (2004). The Caspian Sea. Lake Basin Management Initiative Thematic Paper,
- [5] Alam, M. and O. Maughan (1993). Acute toxicity of selected organophosphorus pesticides to Cyprinus carpio and Barilius vagra. Journal of Environmental Science & Health Part B, 28: 81-89.
- [6] Aydın, R. and K. Köprücü (2005). Acute toxicity of diazinon on the common carp (Cyprinus carpio L.) embryos and larvae. Pesticide Biochemistry and Physiology, 82: 220-225.
- [7] Ayson, F. and T. Lam (1993). Thyroxine injection of female rabbitfish (Siganus guttatus) broodstock: changes in thyroid hormone levels in plasma, eggs, and yolk-sac larvae, and its effect on larval growth and survival. Aquaculture, 109: 83-93.
- [8] Bisson, M. and A. Hontela (2002). Cytotoxic and endocrine-disrupting potential of atrazine, diazinon, endosulfan, and mancozeb in adrenocortical steroidogenic cells of rainbow trout exposed in vitro. Toxicology and Applied Pharmacology, 180: 110-117.
- [9] Boiko, N., O. Vorob'eva, R. Grigor'yan and G. Kornienko (2004). Dynamics of thyroid hormones at early stages of development of the sturgeon Acipenser güldenstadti. Journal of Evolutionary Biochemistry and Physiology, 40: 176-181.
- [10] Brooks, S., C. R. Tyler and J. P. Sumpter (1997). Egg quality in fish: what makes a good egg? Reviews in Fish Biology and Fisheries, 7: 387-416.
- [11] Brown, C., C. Sullivan, H. Bern and W. Dickhoff (1986). Occurrence of thyroid hormones in early developmental stages of teleost fish. 10. Annu. Larval Fish Conf. Miami, Florida, EEUU. 18-23 May 1986.
- [12] Brown, C. L., S. I. Doroshov, M. D. Cochran and H. A. Bern (1989). Enhanced survival in striped bass fingerlings after maternal triiodothyronine treatment. Fish physiology and biochemistry, 7: 295-299.
- [13] Brown, C. L., S. I. Doroshov, J. M. Nunez, C. Hadley, J. Vaneenennaam, R. S. Nishioka and H. A. Bern (1988). Maternal triiodothyronine injections cause increases in swimbladder inflation and

# International Journal of Biological, Life and Agricultural Sciences ISSN: 2415-6612

Vol:11, No:2, 2017

survival rates in larval striped bass, Morone saxatilis. Journal of Experimental Zoology, 248: 168-176.

- [14] Brown, C. L., E. C. Urbinati, W. Zhang, S. B. Brown and M. McComb-Kobza (2014). Maternal Thyroid and Glucocorticoid Hormone Interactions in Larval Fish Development, and Their Applications in Aquaculture. Reviews in Fisheries Science & Aquaculture, 22: 207-220.
- [15] Burkepile, D., M. Moore and M. Holland (2000). Susceptibility of five nontarget organisms to aqueous diazinon exposure. Bulletin of environmental contamination and toxicology, 64: 114-121.
- [16] Cherr, G. N. and W. H. Clark (1982). Fine structure of the envelope and micropyles in the eggs of the white sturgeon, Acipenser transmontanus Richardson. Development, Growth & Differentiation, 24: 341-352.
- [17] Civen, M. and C. B. Brown (1974). The effect of organophosphate insecticides on adrenal corticosterone formation. Pesticide Biochemistry and Physiology, 4: 254-259.
- [18] Cyr, D. G. and J. Eales (1996). Interrelationships between thyroidal and reproductive endocrine systems in fish. Reviews in Fish Biology and Fisheries, 6: 165-200.
- [19] de Jesus, E. G. T. and T. Hirano (1992). Changes in whole body concentrations of cortisol, thyroid hormones, and sex steroids during early development of the chum salmon, Oncorhynchus keta. General and comparative endocrinology, 85: 55-61.
- [20] Dutta, H. and H. Meijer (2003). Sublethal effects of diazinon on the structure of the testis of bluegill, Lepomis macrochirus: a microscopic analysis. Environmental pollution, 125: 355-360.
- [21] Falahatkar, B., S. R. Akhavan and G. Ghaedi (2014). Egg cortisol response to stress at early stages of development in Persian sturgeon Acipenser persicus. Aquaculture international, 22: 215-223.
- [22] Fent, K. and W. Meier (1994). Effects of triphenyltin on fish early life stages. Archives of environmental contamination and toxicology, 27: 224-231.
- [23] Ferrando, M., E. Sancho and E. Andreu-Moliner (1991). Comparative acute toxicities of selected pesticides to Anguilla anguilla. Journal of Environmental Science & Health Part B, 26: 491-498.
- [24] Ghasemzadeh, J., M. Sinaei and M. Bolouki (2015). Biochemical and histological changes in fish, spotted scat (Scatophagus argus) exposed to Diazinon. Bulletin of environmental contamination and toxicology, 94: 164-170.
- [25] Gore, A. C., J. J. Heindel and R. T. Zoeller (2006). Endocrine disruption for endocrinologists (and others). Endocrinology, 147: s1-s3.
- [26] Guideline, O. T. (1992). 203. Fish, Acute Toxicity Test,
- [27] Hajirezaee, S., A. R. Mirvaghefi, H. Farahmand and N. Agh (2016). Effects of diazinon on adaptation to sea-water by the endangered Persian sturgeon, Acipenser persicus, fingerlings. Ecotoxicology and Environmental Safety, 133: 413-423.
- [28] Hamm, J. and D. Hinton (2000). The role of development and duration of exposure to the embryotoxicity of diazinon. Aquatic toxicology, 48: 403-418.
- [29] Hwang, P.-P., S.-M. Wu, J.-H. Lin and L.-S. Wu (1992). Cortisol content of eggs and larvae of teleosts. General and comparative endocrinology, 86: 189-196.
- [30] Katuli, K. K., B. M. Amiri, A. Massarsky and S. Yelghi (2014). Impact of a short-term diazinon exposure on the osmoregulation potentiality of Caspian roach (Rutilus rutilus) fingerlings. Chemosphere, 108: 396-404.
- [31] Khatun, N. and R. Mahanta (2014). A Study on the Effect of Chlorpyrifos (20% EC) on Thyroid Hormones in Freshwater Fish, Heteropneustes fossilis (Bloch.) by using EIA Technique. Science, 2
- [32] Kiabi, B. H., A. Abdoli and M. Naderi (1999). Status of the fish fauna in the South Caspian Basin of Iran. Zoology in the Middle East, 18: 57-65.
- [33] Köprücü, K. and R. Aydın (2004). The toxic effects of pyrethroid deltamethrin on the common carp (Cyprinus carpio L.) embryos and larvae. Pesticide Biochemistry and Physiology, 80: 47-53.
- [34] Kuivila, K. M. and C. G. Foe (1995). Concentrations, transport and biological effects of dormant spray pesticides in the San Francisco Estuary, California. Environmental Toxicology and Chemistry, 14: 1141-1150.
- [35] Li, M., D. P. Bureau, W. A. King and J. F. Leatherland (2010). The actions of in ovo cortisol on egg fertility, embryo development and the expression of growth-related genes in rainbow trout embryos, and the growth performance of juveniles. Molecular reproduction and development, 77: 922-931.
- [36] Mayne, G. J., C. A. Bishop, P. A. Martin, H. J. Boermans and B. Hunter (2005). Thyroid function in nestling tree swallows and eastern bluebirds exposed to non-persistent pesticides and p, p'-DDE in apple orchards of southern Ontario, Canada. Ecotoxicology, 14: 381-396.

- [37] McComb, D. M., J. Gelsleichter, C. A. Manire, R. Brinn and C. L. Brown (2005). Comparative thyroid hormone concentration in maternal serum and yolk of the bonnethead shark (Sphyrna tiburo) from two sites along the coast of Florida. General and comparative endocrinology, 144: 167-173.
- [38] McCormick, M. (1999). Experimental test of the effect of maternal hormones on larval quality of a coral reef fish. Oecologia, 118: 412-422.
- [39] McEnroe, M., N. Purchase and J. J. Cech (1994). Measurements of Thyroid Hormones, T3 and T ', in Juvenile WIIITE Sturgeon over The Annual Cycle.
- [40] Moghim, M., D. Kor, M. Tavakolieshkalak and M. Khoshghalb (2006). Stock status of Persian Sturgeon (Acipenser persicus Borodin, 1897) along the Iranian coast of the Caspian Sea. Journal of Applied Ichthyology, 22: 99-107.
- [41] Nasrabadi, T., G. N. Bidhendi, A. Karbassi, P. Grathwohl and N. Mehrdadi (2011). Impact of major organophosphate pesticides used in agriculture to surface water and sediment quality (Southern Caspian Sea basin, Haraz River). Environmental Earth Sciences, 63: 873-883.
- [42] Osterauer, R. and H.-R. Köhler (2008). Temperature-dependent effects of the pesticides thiacloprid and diazinon on the embryonic development of zebrafish (Danio rerio). Aquatic Toxicology, 86: 485-494.
- [43] Pesando, D., P. Huitorel, V. Dolcini, C. Angelini, P. Guidetti and C. Falugi (2003). Biological targets of neurotoxic pesticides analysed by alteration of developmental events in the Mediterranean sea urchin, Paracentrotus lividus. Marine environmental research, 55: 39-57.
- [44] Pourkazemi, M. (2006). Caspian Sea sturgeon conservation and fisheries: past present and future. Journal of Applied Ichthyology, 22: 12-16.
- [45] Power, D., L. Llewellyn, M. Faustino, M. Nowell, B. T. Björnsson, I. Einarsdottir, A. V. Canario and G. Sweeney (2001). Thyroid hormones in growth and development of fish. Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology, 130: 447-459.
- [46] Rahmanikhah, Z., A. E. Sari, N. Bahramifar and Z. S. Bousjien (2010). Organophosphorous pesticide residues in the surface and ground water in the Southern Coast Watershed of Caspian Sea, Iran. World Applied Sciences Journal, 9: 160-162.
- [47] Raine, J., C. Cameron, M. Vijayan, J. Lamarre and J. Leatherland (2004). The effect of elevated oocyte triiodothyronine content on development of rainbow trout embryos and expression of mRNA encoding for thyroid hormone receptors. Journal of fish biology, 65: 206-226.
- [48] Sadeghi, A. and M. R. Imanpoor (2013). Acute Toxicity of Two Pesticides Diazinon and Deltamethrin on Gambusia. World Journal of Zoology, 8: 303-307.
- [49] Shayeghi, M., M. Khoidel, F. Bageri and M. Abtahi (2008). Azinphos methyl and diazinon residues in rivers of Qara-Su River and Gorgan-rud River in Golestan Province. J Publ Health Health Res Inst, 6: 75-82.
- [50] Shayeghi, M., S. Shahtaheri and M. Selsele (2001). Phosphorous Insecticides Residues in Mazandaran River Waters, Iran (2000). Iranian Journal of Public Health, 30: 115-118.
- [51] Simontacchi, C., E. Negrato, M. Pazzaglia, D. Bertotto, C. Poltronieri and G. Radaelli (2009). Whole-body concentrations of cortisol and sex steroids in white sturgeon (Acipenser transmontanus, Richardson 1836) during early development and stress response. Aquaculture international, 17: 7-14.
- [52] Tagawa, M., M. Tanaka, S. Matsumoto and T. Hirano (1990). Thyroid hormones in eggs of various freshwater, marine and diadromous teleosts and their changes during egg development. Fish Physiology and Biochemistry, 8: 515-520.
- [53] Tanaka, M., J. Tanangonan, M. Tagawa, E. De Jesus, H. Nishida, M. Isaka, R. Kimura and T. Hirano (1995). Development of the pituitary, thyroid and interrenal glands and applications of endocrinology to the improved rearing of marine fish larvae. Aquaculture, 135: 111-126.
- [54] Thangavel, P., K. Sumathiral, S. Karthikeyan and M. Ramaswamy (2005). Endocrine response of the freshwater teleost, Sarotherodon mossambicus (Peters) to dimecron exposure. Chemosphere, 61: 1083-1092.
- [55] Viant, M. R., C. A. Pincetich and R. S. Tjeerdema (2006). Metabolic effects of dinoseb, diazinon and esfenvalerate in eyed eggs and alevins of Chinook salmon (Oncorhynchus tshawytscha) determined by 1 H NMR metabolomics. Aquatic Toxicology, 77: 359-371.
- [56] Vryzas, Z., G. Vassiliou, C. Alexoudis and E. Papadopoulou-Mourkidou (2009). Spatial and temporal distribution of pesticide residues in surface waters in northeastern Greece. Water Research, 43: 1-10.

# International Journal of Biological, Life and Agricultural Sciences ISSN: 2415-6612 Vol:11, No:2, 2017

[57] Welshons, W. V., K. A. Thayer, B. M. Judy, J. A. Taylor, E. M. Curran and F. S. Vom Saal (2003). Large effects from small exposures. I. Mechanisms for endocrine-disrupting chemicals with estrogenic activity. Environmental health perspectives, 111: 994.