Differentiation between Common Tick Species Using Molecular Biology Techniques in Saudi Arabia

Kholoud A. Al-Shammery, Badr El-Sabah A. Fetoh and Ahmed M. Alshammari

Abstract—Protein and Esterase electrophoresis were used to genetically identify two Saudi tick species. Engorged females of the camel tick *Hyalomma dromedarii* (Koch) (Acari: Ixodidae) and the cattle tick *Boophilus annulatus* (Say) (Acari: Ixodidae) ticks collected from infested camels and cattle in the animals resting house at Hail region in KSA were used. The results showed that there are a variation in both of protein and esterase activity levels and a high polymorphism within and between the genera and species of *Hyalomma* and *Boophilus*. In conclusion, the protein and esterase electrophoretic analysis used in the present study could successfully distinguish among tick species, commonly found in Saudi Arabia.

Keywords—Molecular biology, The camel tick *Hyalomma dromedarii*, The cattle tick *Boophilus annulatus*, Ticks.

I. INTRODUCTION

CAMEL ticks *Hyalomma dromedarii* Koch (Acari: Ixodidae) and Cattle ticks *Boophilus annulatus* (Say) (Acari: Ixodidae) are the most serious common parasite of camel and cattle (respectively) in Saudi Arabia. They are disease vectors for different parasites and if uncontrolled, can cause serious losses to the livestock industry [1]. This investigation was aimed to identify these ticks species in Saudi Arabia by using Protein Polyacrylamide Gel Electrophoresis (SDS-PAGE) and Esterase Isozyme Electrophoresis.

The identification of tick species has always been based on morphological key characters of the mouth parts and adjacent structures [2]. These methods cannot be applied to damaged specimens. Recently, protein electrophoresis and molecular genetic studies were introduced to differentiate among the different genera and/or species [3]-[4].

Molecular biology as a new approach helps to classify and control pests in a clear, easy and quick manner [5].

The objectives of this investigation were to assess the possibility of using molecular markers to identify tick species

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based on protein and esterase to estimate the similarity and difference between them.

II. MATERIALS AND METHODS

Collection of Ticks:

Engorged females of the camel tick *Hyalomma dromedarii* (Koch) (Acari: Ixodidae) and the cattle tick *Boophilus annulatus* (Say) (Acari: Ixodidae) ticks collected from infested camels and cattle in the animals resting house at Hail region in KSA.

Molecular Biology Techniques:

Protein Polyacrylamide Gel Electrophoresis (SDS-PAGE):

Preparation for total protein assay was carried out according to the method in [6]. Electrophoresis was carried out as described in [7] using pre-stained high molecular weight standard marker with molecular weight ranged from 200 KDa (KDa = Kilo Dalton) to 6.5 KDa.

After the electrophoresis process the gels were stained with silver stain and distained according to the method in [8]. The stained gels were photographed and examined for the presence and absence of visualized bands.

Esterase Isozyme Electrophoresis:

The same steps were followed for esterase electrophoresis using α – naphthyl propionate as substrate according to [9].

Concentration of protein and esterase bands (Conc. %), relative fragmentation and similarity coefficient (Sim co.) were calculated by following [10] and commonality percentage (Com. %) was calculated according to [11].

Conc. $\% = \underline{O. D. of sample}$ x Conc. Of standard O. D. of standard Where: O. D. = Optical density Rf value = <u>Distance of migrated band</u> Distance of migrated tracked gel

Sim . co. =
$$1 - \underline{NXY}$$

NX+NY

Where:

NXY= The number of common bands in samples X and Y

NX= The number of bands in sample X

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NY= The number of bands in sample Y

Com. % =<u>Number of common bands in samples X and Y</u> Number of total bands of both samples X and Y

III. RESULTS AND DISSCUSSION

Table 1 and Figs (1and 2) showed results of sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) for protein of two species of ticks collected from Saudi Arabia.

The camel tick *Hyalomma dromedarii* showed 16 visualization bands which: 1, 2, 3, 4, 5, 6, 7, 8, 11, 13, 14, 15, 17, 19, 20 and 22. These relative bands ranged between 109.38 KDa and 5.96 KDa, have relative fragmentation (Rf) ranged from 0.012 and 0.917 and concentration varied between 24.44 and 1.23.

TABLE I

QUANTITATIVE PROTEIN PATTERN OF TWO SPECIES OF TICK FROM SAUDI ARABIA REGARDING THE TABLE: (+) PRESENT, (-) ABSENT, MOL.W.= MOLECULAR WEIGHT IN KDA, RF.= RELATIVE FRAGMENTATION, CONC.% = CONCENTRATION PERCENTAGE, SIM. %= 77.14% AND COM. %= 29.03%.

1	+	109.38	0.012	+	108.24	0.167	6.32
2	+	98.42	0.208	+	105.70	0.233	2.13
3	+	95.30	0.250	-	-	-	-
4	+	85.13	0.292	+	98.13	0.292	1.62
5	+	58.21	0.417	+	94.36	0.400	0.74
6	+	40.73	0.442	+	87.18	0.417	1.28
7	+	29.41	0.500	+	39.86	0.500	16.08
8	+	20.80	0.542	-	-	-	-
9	-	-	-	+	27.27	0.583	3.85
10	-	-	-	+	24.08	0.617	1.92
11	+	18.68	0.641	-	-	-	-
12	-	-	-	+	21.06	0.642	2.42
13	+	17.39	0.667	+	19.71	0.667	11.54
14	+	15.75	0.683	+	18.02	0.700	10.36
15	+	14.71	0.708	-	-	-	-
16	-	-	-	+	16.82	0.733	5.14
17	+	12.09	0.733	-	-	-	-
18	-	-	-	+	13.72	0.775	12.80
19	+	9.46	0.792	+	10.83	0.833	20.13
20	+	8.02	0.850	-			
21	-	-	-	+	9.31	0.875	3.44
22	+	5.96	0.917				

TABLE IIQUANTITATIVE ESTERASE PATTERN OF TWO SPECIES OF TICK FROM SAUDIARABIA(+) PRESENT, (-) ABSENT, RF.= RELATIVE FRAGMENTATION, CONC.%= CONCENTRATION PERCENTAGE, SIM. % = 82.93% and COM. %= 50.00%

	The ca	amel tick	c .	The cattle tick			
Band	Hyalomma	a dromed	darii	Boophilus annulatus			
number	Band			Band			
	occurrence	Rf.	Con.	occurrence	Rf.	Con.	
			%			%	
1	+	0.19	1	+	0.19	1	
2	+	0.23	2	+	0.23	2	
3	+	0.40	3	+	0.40	3	
4	+	0.46	4	+	0.46	4	
5	+	0.50	5	+	0.50	5	
6	+	0.69	6	+	0.69	6	
7	+	0.72	7	+	0.72	7	
8	+	0.93	8	+	0.93	8	

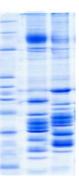


Fig. 1. SDS- polyacrylamide gel zymogram of denatured protein patterns in two species of ticks attacking domestic animals in KSA.1 and 2 represent lanes of tick samples, where 1 = the camel tick *Hyalomma dromedarii* and 2 = the cattle tick *Boophilus annulatus* and lane M represents the known molecular size marker.

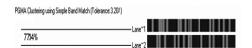


Fig. 2. Similarity relationships among protein bands in the camel tick *Hyalomma dromedarii* and the cattle tick *Boophilus annulatus* (Say) in KSA.

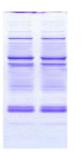


Fig. 3. Polyacrylamide gel zymogram of esterase isozyme patterns in two species of ticks in KSA. 1 = the camel tick *Boophilus annulatus* and 2 = the cattle tick *Boophilus microplus*.

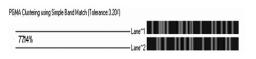


Fig. 4. Similarity relationships among esterase bands in the camel tick *Hyalomma dromedarii* and the cattle tick *Boophilus annulatus* in KSA.

On the other hands, the cattle tick *Boophilus annulatus* has 15 bands : 1, 2, 4, 5, 6, 7, 9, 10, 12, 13, 14, 16, 18, 19, 21. These bands were located between 108.24 KDa and 9.31 KDa, have Rf values ranged from 0.167 and 0.875 and concentration varied from 6.32 and 3.44. The common reactive bands between the camel tick *Hyalomma dromedarii* and the cattle tick *Boophilus annulatus* were nine bands : 1, 2, 4, 5, 6, 7, 13, 14 and 19, with molecular weight ranged from 109.38 KDa and 9.46 KDa. Similarity % was 77.14% and the commonality % was 29.03%.

Data in Table 2 and figs (3 and 4) show esterase profile pattern of two different species of ticks in Saudi Arabia, the first species is the camel tick *Hyalomma dromedarii* and the second species is the cattle tick *Boophilus annulatus*. Both of species have eight different esterase bands , with Rf values ranged from 0.19 to 0.93 , respectively, in both two tick species and concentration ranged from 8.78 to 36.14 in the camel tick and from 8.59 to 35.04 in the cattle tick, respectively. Similarity % in esterase bands of ticks species was 82.93% and the commonality % was 50.00%.

In the same trend [13] detected fifteen negatively charged protein bands were found by acrylamide-gel electrophoresis to be present in the whole blood of the cattle tick *Boophilus microplus* in Australia. The bands were further characterized into glycoproteins, haemoproteins, esterases, phosphatases, and an aminopeptidase. Reference [5] utilized SDS-PAGE and esterase profile patterns to discrimination between two species of fruit flies and in [14] used SDS-PAGE to detect midgut antigens of *Hyalomma anatolicum anatolicum* tick.

Isozyme was used for species identification of acarines. The most commonly studied enzymatic system in mites is that of the esterases [15].

In ticks, several enzymatic systems can be resolved from an individual. However, diverse studies have reported low polymorphism of the resolved loci [13] – [16]. Furthermore, Reference [17] used Esterase and RAPD-PCR analysis to differentiate among four species of ticks in Egypt, these tick species were: *Argas hermanni* (Audouin), *Argas persicus* (Oken), *Hyalomma dromedarii* (Koch), *Hyalomma anatolicum excavatum* (Koch).

REFERENCES

- [1] J. R. Mohler, "Tick Fever," *Agri. Farmer's Bull.*, 1990, vol. 1625, pp 1-30
- [2] K. L. Poucher, H. J. Hutcheson, J. E. Keirans, L. A. Durden, and W. C. Black, "IV. Molecular genetic key for the identification of 17 *Ixodes* species of the United States (Acari: Ixodidae): A methods model", J. Parasito., 1999, vol. 85(4), pp. 623-629.
- [3] K. M. El Kammah, and M. A. Sayed, "Biochemical fingerprints of egg and salivary gland proteins characterising four common tick genera in Egypt", 1999.
- [4] K. L. Poucher, H. J. Hutcheson, J. E. Keirans, L. A. Durden, and W. C. Black, "Molecular genetic key for the identification of 17 *Ixodes* species of the United States (Acari: Ixodidae): a methods model". *J. Parasitol.*, vol. 85, 1999, pp. 623-629.
- [5] B. E. A. Fetoh, "Molecular and modern taxonomy of *Dacus* spp. (Diptera: Tephritidae) attacking vegetables in Egypt", *J. Egypt. Ger. Soc. Zool.*, 2006, vol. 48E, pp. 1-21.
- [6] O. H. Lowry, C. S. Rosebrough, and R. J. Randall, "Protein measurement with the folin phenol reagent', J. Biolo. Chem., 1951, vol. 193, pp. 265-275.
- [7] U. K. Laemmli, "Cleavage of structural proteins during assembly of head bacteriophage T₄", *Nature*, 1970, vol. 227, pp. 680- 685.
- [8] P. J. Hitchcock, and T. M. Brown, "Morphological heterogeneity among Salmomella lipopolysaccaride types in Silver – stained poly acrylamide gel", J. Bact., 1983, vol. 154, pp. 269-277.
- [9] Sims, M. "Methods for detection of enzymatic activity after electrophoresis on polyacrylamide gel in *Drosophila* species" *Nature*, 1965, vol. 207(14), pp. 757-758.
- [10] M. Nei, and W.H. Li, "Mathematical model for studying genetic variation in terms of restriction endonucleases", *Proc. Natl. Acad. Sci.* (USA), 1979, vol. 76, pp. 5269-5273.
- [11] Y. Yorozu, M. Hirano, K. Oka, and Y. Tagawa, "Electron spectroscopy studies on magneto-optical media and plastic substrate interfaces(Translation Journals style)," *IEEE Transl. J. Magn.Jpn.*, vol. 2, Aug. 1987, pp. 740–741 (*Dig. 9th Annu. Conf. Magnetics* Japan, 1982, p. 301).
- [12] D.S. Hayer, and D. McInnis, "Resolution of populations of the Mediterranean fruit fly *Ceratitis capitata* at DNA level using random primers)," *Genome*, 1994, vol. 37, pp. 244-248.
- [13] Tatchell, R.J. Electrophoretic studies on the proteins of the haemolymph, saliva, and eggs of the cattle tick, *Boophilus microplus*. *Insect Biochem.*, 1971, vol.1, no. 1, pp. 45-55.
- [14] C. Delaye, L. Beati, A. Aeschimann, F. Renaud, and T. de Meeus, "Population genetic structure of *Ixodes ricinus* in Switzerland from allozymic data: no evidence of divergence between nearby sites", Int. J. Parasitol., 1997, vol. 27, pp. 769-773.
- [15] Gh. Norouzi, R. Hashemitabar, and G. R. Razmi, "Detection of midgut antigens of *Hyalomma anatolicum anatolicum* tick using SDS-PAGE and Western Blot", Iranian Journal of Veterinary Research, Shiraz University, 2007, vol. 8 (2), pp. 166-169.
- [16] A. M. Avanzati, M. Baratti, and F. Bernini," Molecular and morphological differentiation between steganacarid mites (Acari:Oribatida)", *Biol. J. Linn. Soc.*, 1994, vol. 52, pp. 25-340.
 [17] D. E. Kain, F. A. Sperling, and R.S. Lane, "Population genetic
- [17] D. E. Kain, F. A. Sperling, and R.S. Lane, "Population genetic structure of *Ixodes pacificus* (Acari: Ixodidae) using allozymes", J. Med. Entomol., 1997, vol. 34, pp. 441-450.
- [18] Z.A. El-Fiky, M.A. Sayed and K.M. El Kammah, "Esterase and RAPD-PCR analysis of common tick species in Egypt", *Arab J. Biotech.* 2003, vol. 6, no.1, pp. 39-48.